

BD Rhapsody™ Single-Cell Analysis System Instrument User Guide

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Becton, Dickinson and Company
BD Biosciences
2350 Qume Drive
San Jose, CA 95131 USA
Tel 1.877.232.8995

bdbiosciences.com
ResearchApplications@bd.com

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History

Revision	Date	Change made
Doc ID: 214062 Rev. 1.0	07/2018	Initial release
Doc ID: 214062 Rev. 2.0 23-21336-00	02/2019	Added information about the BD™ Mouse Immune Single-Cell Multiplexing Kit.

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Introduction

- About this guide (page 8)
- Intended use (page 8)
- Safety symbols (page 10)
- Safety data sheets (page 12)
- Instrument technical support (page 13)

About this guide

Introduction

This guide provides instructions on operating the BD Rhapsody™ Scanner, the BD Rhapsody™ Express instrument and supporting materials.

To use the BD Rhapsody™ Single-Cell Analysis system without the BD Rhapsody Scanner, see the *BD Rhapsody™ Express Single-Cell Analysis System Instrument User Guide* (Doc ID: 214063).

Genomics technical publications are available for download from the BD Genomics Resource Library at bd.com/genomics-resources.

Intended use

Intended use

The BD Rhapsody Single-Cell Analysis system is intended for the preparation of single cell sequencing libraries. The BD Rhapsody Scanner uses brightfield and dual band fluorescence to count cells and measure cell viability. Proprietary BD™ Molecular Indexing technology is used to count individual mRNA molecules. Cells are entirely contained in the BD Rhapsody™ Cartridge, which is a single-use consumable.

The system is intended for use by professional scientific users, such as technicians and laboratory personnel, who are trained in the operation of the BD Rhapsody Express instrument and BD Rhapsody Scanner.

For Research Use Only. Not for use in diagnostic or therapeutic procedures.

For more information on the purpose of the instruments, see the [BD Rhapsody™ Express instrument overview \(page 18\)](#) and the [BD Rhapsody™ Scanner overview \(page 20\)](#).

Restrictions

Any use of the BD Rhapsody Single-Cell Analysis system other than the procedures described in this user guide might result in damage to the instruments, loss of reagents or samples, or personal injury.

BD denies any responsibility for damage caused by the following:

- Any use of a BD Rhapsody™ instrument that does not comply with the procedures described in any guide used with the BD Rhapsody Single-Cell Analysis system
- Unauthorized alterations or adjustments to instrument hardware or software
- Any use of an instrument that violates locally applicable laws, rules, or regulations
- Evidence of any deviation from intended use voids the BD Rhapsody instrument warranty

Disclaimer

The instrument, external components, software, and consumables in the BD Rhapsody Single-Cell Analysis system are provided for research purposes only. BD disclaims all express and implied warranties, including, but not limited to, merchantability and fitness for use for a particular purpose.

Safety symbols



Introduction

This topic describes the safety symbols used in this guide.






For safety and limitations, see the *BD Rhapsody™ Express Instrument and BD Rhapsody™ Scanner Safety and Limitations Guide* (Doc ID: 42061).

Safety symbols

The following table lists the safety symbols used in this guide to alert you to potential hazards.

Symbol	Meaning
	Caution. Indicates the need for the user to consult the instructions for use for important cautionary information, such as warnings and precautions that cannot, for a variety of reasons, be presented on the device itself.
	Biological hazard. All surfaces that come in contact with biological specimens can transmit potentially fatal disease. Use universal precautions when cleaning surfaces. Wear suitable protective clothing, eyewear, and gloves.

Other symbols

Symbol	Meaning
 The symbol consists of the letters "REF" in a bold, sans-serif font, enclosed within a rectangular border.	Part number
 The symbol consists of the letters "LOT" in a bold, sans-serif font, enclosed within a rectangular border.	Lot number
 A simple line drawing of a thermometer, showing a bulb at the bottom and a vertical scale with a horizontal line indicating a temperature level.	Storage temperature range
 A simple line drawing of an hourglass, consisting of two inverted cones joined at their tips.	Expiration date
 A simple line drawing of an open book with a lowercase letter 'i' on the right page.	Consult instructions for use

Safety data sheets

Introduction

This topic describes how to obtain safety data sheets (SDSs).

Obtaining SDSs

Before handling chemicals, read and understand the SDSs. To obtain SDSs for chemicals ordered from BD Biosciences, go to regdocs.bd.com, or contact BD Biosciences technical support at researchapplications@bd.com.

Instrument technical support

Introduction This topic describes how to get technical support for the BD Rhapsody Express instrument and BD Rhapsody Scanner.

Contacting technical support If technical assistance is required, contact BD Biosciences technical support at researchapplications@bd.com or 1.877.232.8995, prompt 2, 2. You can contact technical support in Europe at help.biosciences@europe.bd.com or at these telephone numbers:

Location	Telephone number	Location	Telephone number
Worldwide	+32 2 40 09 895	—	
Austria	01 92 80 465	Netherlands	010 71 14 800
Belgium	02 40 17 093	Norway	800 18 530
Denmark	80 88 21 93	Portugal	800 86 01 76
Finland	800 11 63 17	South Africa	0800 98 10 08
France	01 70 70 81 93	Spain	91 41 46 250
Germany	069 22 22 25 60	Sweden	08 50 69 21 54
Greece	00800 12 75 06	Switzerland	044 58 04 373
Italy	02 36 00 36 85	United Kingdom	0207 07 53 226

Before contacting BD Biosciences, have the following information available:

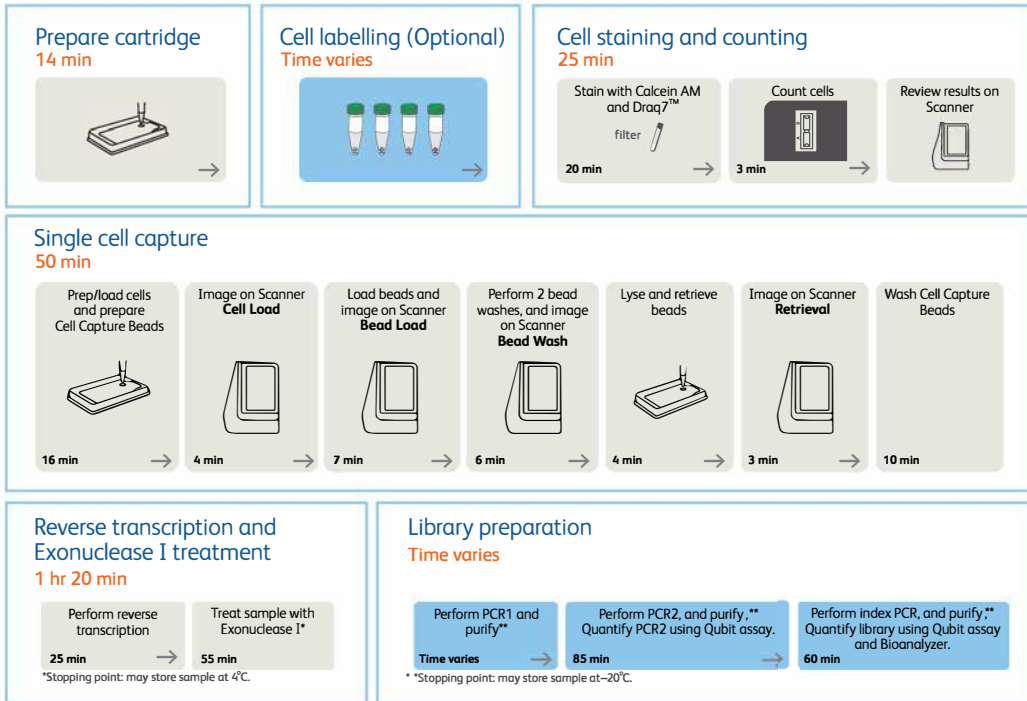
- Product name, part number, and serial number or lot number
 - Any error messages
 - Details of recent system performance
 - For the BD Rhapsody Scanner, the version of the software that you are using
-

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Getting Started

- [BD Rhapsody™ targeted library workflow \(page 16\)](#)
- [Site requirements \(page 17\)](#)
- [Thermomixer settings \(page 17\)](#)
- [Thermal cycler setup \(page 17\)](#)
- [Pipette Settings \(page 18\)](#)
- [BD Rhapsody™ Express instrument overview \(page 18\)](#)
- [BD Rhapsody™ Scanner overview \(page 20\)](#)
- [Quick guide to the BD Rhapsody Scanner software \(page 23\)](#)
- [Best practices \(page 27\)](#)

BD Rhapsody™ targeted library workflow



To perform the workflow, follow the *Single Cell Analysis Workflow with BD Rhapsody™ Systems* (Doc ID: 220524).

Note: For use of two cartridges, see [Workflow with two BD Rhapsody™ Cartridges](#) (page 129).

Site requirements

Workspace designation

Dedicate two isolated workspaces in the laboratory to run high-sensitivity, single cell sequencing experiments:

- Pre-amplification workspace
- Post-amplification workspace

For detailed site requirements and technical specifications, see the *BD Rhapsody™ Express Instrument and BD Rhapsody™ Scanner Site Preparation Guide* (Doc ID: 47391) and [BD Rhapsody™ Scanner metrics \(page 119\)](#).

For installation of the BD Rhapsody Express instrument and the BD Rhapsody Scanner, see the *BD Rhapsody™ Single-Cell Analysis System Installation and Maintenance Guide* (Doc ID: 43084).

Genomics technical publications are available for download from the BD Genomics Resource Library at bd.com/genomics-resources.

Thermomixer settings

Settings

Depending on the protocol, set the thermomixer from 37°C to 80°C and 0–1,200 rpm.

Thermal cycler setup

Recommendations

- Use a properly calibrated thermal cycler for 0.2 mL tubes with a maximum reaction volume of 50 μ L.
- Use a heated lid set to $\geq 95^{\circ}\text{C}$.
- Do not use fast cycling mode.

- For specific instrument operation, follow the instructions provided by the manufacturer.
-

Pipette Settings

Pipette programs BD Rhapsody™ P1200M and P5000M pipettes are provided pre-programmed for use during single cell mRNA capture from the BD Rhapsody Cartridge. Do not change the settings, but confirm them before use.

Pipette	Mode
P1200M	Prime/Treat
P1200M	Cell Load
P1200M	Bead Load
P1200M	Wash
P1200M	Lysis
P5000M	Retrieval

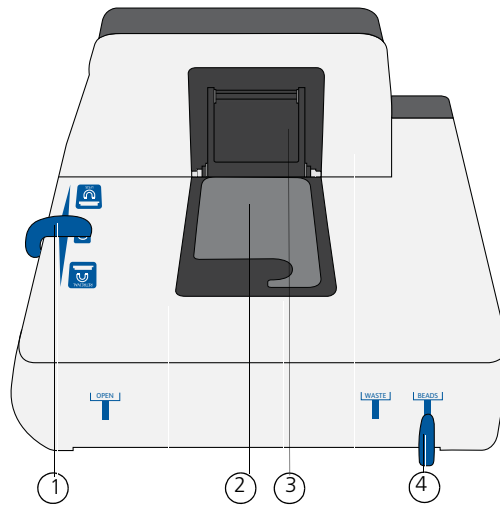
BD Rhapsody™ Express instrument overview

Introduction The BD Rhapsody Cartridge requires use of the BD Rhapsody Express instrument. The station is used to load reagents, cells, and Cell Capture Beads into the cartridge for bead capture and retrieval of single cell mRNA.

Safety For safety and limitations of the BD Rhapsody Express instrument, see the *BD Rhapsody™ Express Instrument and BD Rhapsody™ Scanner Safety and Limitations Guide* (Doc ID: 42061).

Components

The following figure shows the main components of the BD Rhapsody Express instrument for operation. For maintenance of the Express instrument, see the *BD Rhapsody™ Single-Cell Analysis System Installation and Maintenance Guide* (Doc ID: 43084).



No.	Component
1	Left slider to position Retrieval (top) and Lysis (bottom) magnets. Slider shown in 0 (neutral) position: no magnets applied to BD Rhapsody Cartridge
2	Cartridge tray to install the BD Rhapsody Cartridge
3	Retrieval (top) magnet in up position
4	Front slider to position Waste Collection Container (WASTE) and 5 mL LoBind Tube for bead retrieval (BEADS) and for Waste Collection Container and 5 mL LoBind Tube access (OPEN)

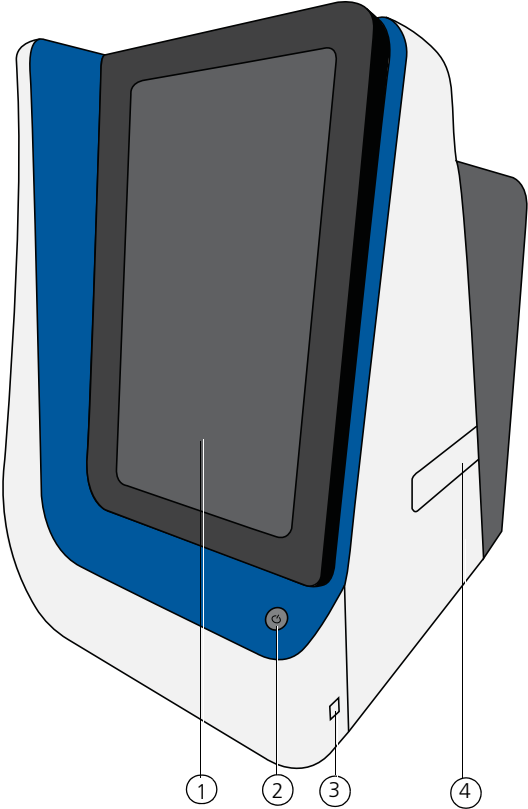
BD Rhapsody™ Scanner overview

Introduction The BD Rhapsody Scanner is used to count cells by brightfield and dual band fluorescence imaging and calculate volumes of cells and Sample Buffer needed to prepare single cell suspensions.

Safety For safety and limitations of the scanner, see the *BD Rhapsody™ Express Instrument and BD Rhapsody™ Scanner Safety and Limitations Guide* (Doc ID: 42061).

Installation and maintenance See the *BD Rhapsody™ Single-Cell Analysis System Installation and Maintenance Guide* (Doc ID: 43084) to review critical installation and periodic maintenance procedures on the scanner.

Components The following figure shows the main components of the scanner for operation:



No.	Front component
1	Scanner touchscreen
2	Scanner front power button and standby switch. Lit green: ON. Flashing: PC asleep.
3	USB 3.0 port
4	Cartridge loading door where a tray extends and retracts

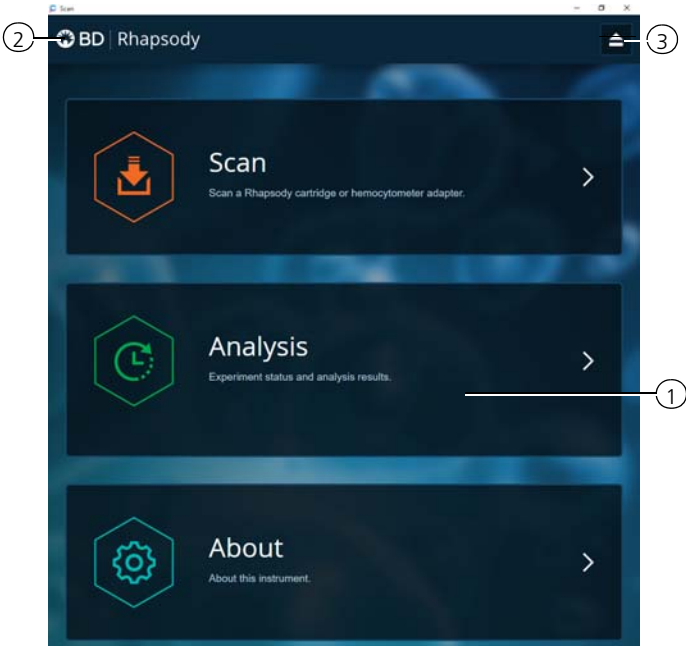


No.	Rear component
1	Master power switch
2	Fuse holder
3	24 VDC power input
4	USB 2.0 ports (2) ^a
5	Ethernet port

- a. The USB 2.0 ports on the rear of the instrument are primarily used by BD Biosciences technical support.

Quick guide to the BD Rhapsody Scanner software

Navigation



No.	Description
1	Tap an application (app) to navigate between scan, analysis, and about screens.
2	Tap Rhapsody on any screen to return to the main screen.
3	(Optional) Tap the eject icon to eject the tray from the sample loading door of the scanner. The scanner automatically ejects the tray.

The screenshot shows the 'Samples Calculator' interface in the BD Rhapsody software. The interface includes a top navigation bar with 'Scan', 'Experiment', and 'About' buttons. The main area is titled 'Samples Calculator' and contains several input fields and a table.

Callout 4 points to the 'Select Samples' button at the top left of the calculator area.

Callout 5 points to the 'Desired total volume' input field, which is set to 650 μl .

Callout 6 points to the 'Relative Amounts' column in the sample table, specifically to the '1' value for the Jurkat sample.

Callout 7 points to the 'Stock Volume' column in the 'Results' section, specifically to the '17.2 μl ' value for the Ramos sample.

At the bottom of the calculator area, there is a green 'Save' button, which is also pointed to by callout 4.

Sample	Time	Concentration (cells/ μl)	Viable Cells (%)	Relative Amounts
Jurkat	2018-01-31 10:01:43.07	423.64	92.24	- 1 +
Ramos	2018-01-31 10:02:39.54	375.99	92.53	- 1 +

Sample	Stock Volume
Jurkat	15.3 μl
Ramos	17.2 μl
Buffer volume	617.5 μl
Loading cell concentration	19.9 cells/ μl
Estimated cell doublet rate	2.4%

No.	Description
4	Tap Select Samples to display the window for selecting samples. Tap Calculate to calculate volumes for preparing cell suspensions. Tap Save on the Calculate tab to display saved calculations.
5	Enter the total volume and desired number of captured cells to prepare the cell suspension for loading into the BD Rhapsody Cartridge.
6	(Optional) Enter the relative amounts of samples that comprise the cell suspension. The default is 1:1.
7	Obtain the volumes of buffer and stock cells required to prepare the cell suspension for cartridge loading.

**BD Rhapsody
Scanner software
workflows**

Scan application workflow

Step	Screen
1	Main menu
2	Hemocytometer/cartridge placement on tray
3	Scan
4	Cartridge removal

Analysis application workflow

Step	Screen
1	Main menu
2	Experiment details
3	Prepare sample
4	Select sample
5	Calculate cell suspension volumes
6	Save calculation
7	Analysis metrics

Best practices

Good laboratory practices

- Calibrate and service pipettes every 12 months to ensure accurate sample volume transfer at each step. To clean and calibrate the pipettes, see the *BD Rhapsody™ Single-Cell Analysis System Installation and Maintenance Guide* (Doc ID: 43084).
 - Unless otherwise specified, thaw reagents on ice. Store reagents at their specified storage conditions.
 - Collect small volumes by briefly centrifuging samples. Brief or pulse centrifugation is <1 second.
 - Gently vortex solutions containing enzymes. Minimize vortex duration and keep the vortex speed low. Do not vortex solutions containing Cell Capture Beads.
 - Gently pipet cells to avoid cell stress or death.
 - Work in designated pre- or post-amplification workspaces according to the protocol.
 - Prepare reagent mixes in pre-amplification workspaces, and conduct amplification in post-amplification workspaces.
 - Wear suitable protective clothing, eyewear, and gloves.
-

RNase-free technique

Prevent the introduction of exogenous RNases into samples during processing:

- Use low-retention, RNase-free pipette tips and low-binding reaction tubes when required for certain steps to prevent absorption to plastic surfaces and minimize bead loss.
 - Wear disposable gloves, and change them frequently.
 - Never reuse tips or tubes.
 - Keep tip boxes, reagent containers, and sample tubes closed when not in use.
 - Always maintain a clean laboratory bench, and if necessary, wipe work surface with a solution of 10% bleach.
-

Sterility

- Clean cell culture surfaces in the laminar flow hood with 70% ethyl alcohol, and appropriately sterilize the surfaces.
 - Use sterile serological pipettes to aseptically transfer media and cells.
 - Place a flask in a cell culture hood one at a time to prevent cross-cell contamination.
-

Cell Capture Beads

- Keep the Cell Capture Beads on ice before use.
Do not freeze Cell Capture Beads.
 - For maximum recovery, do not vortex samples containing Cell Capture Beads.
 - Gently mix suspensions with Cell Capture Beads by pipette only.
 - Use low retention tips and LoBind tubes when handling Cell Capture Beads.
-

BD Rhapsody Express instrument

- The Express instrument contains strong magnets. Avoid having metal pieces close to the station.
 - Wipe the Express instrument with 70% ethyl alcohol wipes after each use.
-

BD Rhapsody Cartridge

- Avoid pipetting bubbles into the cartridge. Before adding fluid to the cartridge, ensure that the pipette tip does not contain air.
 - To ensure an air-tight seal with the BD Rhapsody™ P1200M and P5000M pipettes, hold the pipette with one hand, and slightly twist the pipette to firmly seat a pipette tip on the pipette shaft.
 - Cells need to be prepared as close to cell loading in the cartridge as possible. Begin cell preparation during or after the prime or substrate treatment steps, and leave the cartridge in Cartridge Wash Buffer 2 (Cat. No. 650000061) until ready to proceed with cell loading.
-

**BD Rhapsody
Scanner software**

For easy access to analysis metrics, place the Rhapsody Data folder into the Quick Access folder:

1. Open a file explorer window.
 2. Enter **Public Documents** in the location box.
 3. Drag the **Rhapsody Data** folder to **Quick Access**.
 4. Open the **Rhapsody Data** folder.
 5. Open the folder for the appropriate experiment.
 6. Open the .csv files with Microsoft® Excel.
-

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Product information

- Required kits and storage conditions (page 32)
- Suggested kits (page 42)
- Reagents (page 45)
- Consumables (page 48)
- Equipment (page 51)
- Software (page 55)

Required kits and storage conditions

Introduction

The BD Rhapsody™ Targeted mRNA and AbSeq Reagent Kit—4 Pack (Cat. N. 633771) consists of four boxes:

- BD Rhapsody™ Cartridge Reagent Kit
- BD Rhapsody™ Cartridge Kit
- BD Rhapsody™ cDNA Kit
- BD Rhapsody™ Targeted mRNA and AbSeq Amplification Kit

The BD Rhapsody Targeted mRNA and AbSeq Amplification Kit (Cat. No. 633774) contains sufficient amplification reagents to prepare four libraries of each type, including targeted mRNA, Sample Tag, and BD™ AbSeq libraries. Sub-sampling of the Exonuclease I-treated Cell Capture Beads might require purchasing additional BD Rhapsody Targeted mRNA and AbSeq Amplification kits.

To perform sample multiplexing with the BD Rhapsody™ system, one of the following additional kits are required:

- BD™ Human Single-Cell Multiplexing Kit (Cat. No. 633781)
- BD™ Mouse Immune Single-Cell Multiplexing Kit (Cat. No. 633793)

To perform antibody-oligonucleotide labelling, use additional these components:

- BD™ AbSeq Ab-Oligos (antibody-oligonucleotides)
- BD Pharmingen™ Stain Buffer (FBS)
- (Optional) BD Pharmingen™ Human BD Fc Block™, for use with myeloid and B lymphocyte-containing samples

Product information on the kits and reagents are in this chapter.

**BD Rhapsody
Targeted mRNA
and AbSeq Reagent
Kit—4 Pack (Cat.
No. 633771)**

- Store the four kit boxes at the specified storage temperatures. Use only non-frost free freezers for reagent storage.
- Keep the reagents on ice unless instructed otherwise.
- The BD Rhapsody™ Cartridge is single-use only.
- Limit preparation of mixes to $\leq 20\%$ overage.

Box	Component	Cap color	Quantity	Volume per unit	Storage
BD Rhapsody Cartridge Reagent Kit (Cat. No. 633731)	Cartridge Wash Buffer 1	Neutral	1 bottle	7 mL	2°C to 8°C
	Cartridge Wash Buffer 2	Neutral	1 bottle	4 mL	
	Sample Buffer	Neutral	1 bottle	28 mL	
	Lysis Buffer	Neutral	4 bottles	15 mL	
	Bead Wash Buffer	Neutral	1 bottle	10 mL	
	1 M DTT	White	1 vial	400 μ L	
	Cell Capture Beads	Brown	4 vials	2 mL	
Waste Collection Container	Neutral	4 each	—		

Box	Component	Quantity	Storage
BD Rhapsody Cartridge Kit (Cat. No. 633733)	BD Rhapsody Cartridge	4 each	15°C to 25°C

Box	Component	Cap color	Quantity	Volume per unit	Storage
BD Rhapsody cDNA Kit (Cat. No. 633773)	Nuclease-Free Water	Neutral	2 vials	1 mL	-25°C to -15°C
	RT Buffer	Orange	1 vial	200 µL	
	RT 0.1M DTT	Orange	1 vial	50 µL	
	Reverse Transcriptase	Orange	1 vial	50 µL	
	dNTP	Orange	1 vial	100 µL	
	RNase Inhibitor	Orange	1 vial	50 µL	
	Bead RT/PCR Enhancer	Black	1 vial	70 µL	
	10X Exonuclease I Buffer	Yellow	1 vial	100 µL	
	Exonuclease I	Yellow	1 vial	50 µL	
	Bead Resuspension Buffer	Black	1 vial	1 mL	

Box	Component	Cap color	Quantity	Volume per unit	Storage
BD Rhapsody Targeted mRNA and AbSeq Amplification Kit (Cat. No. 633774)	Nuclease-Free Water	Neutral	1 vial	1 mL	-25°C to -15°C
	Bead RT/PCR Enhancer	Black	1 vial	70 µL	
	PCR MasterMix	White	1 vial	1.2 mL	
	Elution Buffer	Pink	1 vial	1.8 mL	
	Universal Oligo	White	1 vial	130 µL	
	Library Forward Primer	Red	1 vial	40 µL	
	Library Reverse Primer 1	Red	1 vial	20 µL	
	Library Reverse Primer 2	Red	1 vial	20 µL	
	Library Reverse Primer 3	Red	1 vial	20 µL	
	Library Reverse Primer 4	Red	1 vial	20 µL	
	Bead Resuspension Buffer	Black	1 vial	1 mL	
	Sample Tag PCR1 Primer	Purple	1 vial	20 µL	
	Sample Tag PCR2 Primer	Purple	1 vial	20 µL	
	BD AbSeq Primer	Green	1 vial	70 µL	

Kit	Components	Quantity	Volume per unit	Storage
BD Human Single-Cell Multiplexing Kit (Cat. No. 633781)	Sample Tag 1— Human	1 vial	20 µL	2°C to 8°C
	Sample Tag 2— Human	1 vial	20 µL	
	Sample Tag 3— Human	1 vial	20 µL	
	Sample Tag 4— Human	1 vial	20 µL	
	Sample Tag 5— Human	1 vial	20 µL	
	Sample Tag 6— Human	1 vial	20 µL	
	Sample Tag 7— Human	1 vial	20 µL	
	Sample Tag 8— Human	1 vial	20 µL	
	Sample Tag 9— Human	1 vial	20 µL	
	Sample Tag 10— Human	1 vial	20 µL	
	Sample Tag 11— Human	1 vial	20 µL	
	Sample Tag 12— Human	1 vial	20 µL	

Kit	Components	Quantity	Volume per unit	Storage
BD Mouse Immune Single-Cell Multiplexing Kit (Cat. No. 633793)	Sample Tag 1— Mouse Immune	1 vial	20 µL	2°C to 8°C
	Sample Tag 2— Mouse Immune	1 vial	20 µL	
	Sample Tag 3— Mouse Immune	1 vial	20 µL	
	Sample Tag 4— Mouse Immune	1 vial	20 µL	
	Sample Tag 5— Mouse Immune	1 vial	20 µL	
	Sample Tag 6— Mouse Immune	1 vial	20 µL	
	Sample Tag 7— Mouse Immune	1 vial	20 µL	
	Sample Tag 8— Mouse Immune	1 vial	20 µL	
	Sample Tag 9— Mouse Immune	1 vial	20 µL	
	Sample Tag 10— Mouse Immune	1 vial	20 µL	
	Sample Tag 11— Mouse Immune	1 vial	20 µL	
	Sample Tag 12— Mouse Immune	1 vial	20 µL	

**BD Rhapsody™
targeted primer
panels**

Each panel includes a set of primers designed to target human (Hs) or mouse (Mm) genes. Each panel contains sufficient primers to prepare four libraries.

A supplement to a panel can be designed based on individual needs. The BD Rhapsody™ Panel Supplement contains primer pairs for additional targeted sequencing of selected genes. For more information on BD Rhapsody Panel Supplements, contact BD Biosciences technical support at researchapplications@bd.com and see [BD Rhapsody™ Panel Supplement \(Cat. No. 633742\) \(page 43\)](#).

Panel	Component	Cap color	Quantity	Vol. (µL)	Storage
BD Rhapsody™ Immune Response Panel Hs (Cat. No. 633750)	PCR1 Primers-Immune Response Panel Hs (tube label: PCR1-Immune Res. Hs)	Blue	1 vial	210	-25°C to -15°C
	PCR2 Primers-Immune Response Panel Hs (tube label: PCR2-Immune Res. Hs)	Blue	1 vial	50	
BD Rhapsody™ T Cell Expression Panel Hs (Cat. No. 633751)	PCR1 Primers-T Cell Expression Panel Hs (tube label: PCR1-T Cell Expression Hs)	Blue	1 vial	210	-25°C to -15°C
	PCR2 Primers-T Cell Expression Panel Hs (tube label: PCR2-T Cell Expression Hs)	Blue	1 vial	50	
BD Rhapsody™ Onco-BC Panel Hs (Cat. No. 633752)	PCR1 Primers-Onco-BC Panel Hs (tube label: PCR1-Onco-BC Hs)	Blue	1 vial	210	-25°C to -15°C
	PCR2 Primers-Onco-BC Panel Hs (tube label: PCR2-Onco-BC Hs)	Blue	1 vial	50	

Panel (continued)	Component	Cap color	Quantity	Vol. (µL)	Storage
BD Rhapsody™ Immune Response Panel Mm (Cat. No. 633753)	PCR1 Primers-Immune Response Panel Mm (tube label: PCR1-Immune Res. Mm)	Blue	1 vial	210	-25°C to -15°C
	PCR2 Primers-Immune Response Panel Mm (tube label: PCR2-Immune Res. Mm)	Blue	1 vial	50	

BD Rhapsody™ Targeted mRNA and AbSeq Training Kit—4 Pack (Cat. No. 633772)

For detailed information on the kit components, see [BD Rhapsody Targeted mRNA and AbSeq Reagent Kit—4 Pack \(Cat. No. 633771\)](#) (page 33).

Component	Quantity	Storage
BD Rhapsody Cartridge Reagent Kit (Cat. No. 633731)	1 kit	2°C to 8°C
BD Rhapsody Cartridge Kit (Cat. No. 633733)	1 kit	15°C to 25°C
BD Rhapsody cDNA Kit (Cat. No. 633773)	1 kit	-25°C to -15°C
BD Rhapsody Targeted mRNA and AbSeq Amplification Kit (Cat. No. 633774)	1 kit	
BD Rhapsody Immune Response Panel Hs (Cat. No. 633750)	1 kit	
BD Rhapsody™ Training Cells (Cat. No. 633741) <ul style="list-style-type: none"> • Jurkat • Ramos 	<ul style="list-style-type: none"> • 2 vials, 1.0 mL each • 2 vials, 1.0 mL each 	Liquid nitrogen

**Required kits from
other vendors**

Kit	Supplier	Catalog no.
Qubit™ dsDNA HS Assay Kit	Thermo Fisher Scientific	Q32851

Suggested kits

BD Rhapsody™ Custom Panel

The BD Rhapsody Custom Panel contains a maximum of 500 custom primers. To order a BD Rhapsody Custom Panel, contact BD Biosciences at researchapplications@bd.com.

Panel	Component	Cap color	Quantity	Volume	Storage
BD Rhapsody Custom Panel (U.S. only: Cat. No. 633743)	BD Rhapsody™ 10X PCR1 Custom ID: xxxxxx ^a	Blue	1 vial	1.4 mL ^b	-25°C to -15°C
	BD Rhapsody™ 10X PCR2 Custom ID: xxxxxx ^a	Blue	1 vial	350 µL ^b	
	IDTE pH 8.0	Blue	1 vial	2.0 mL	
BD Rhapsody Custom Panel 2–99 genes (Outside U.S.: Cat. No. 633777)	BD Rhapsody™ 10X PCR1 Custom ID: xxxxxx ^a	Blue	1 vial	1.4 mL ^b	-25°C to -15°C
	BD Rhapsody™ 10X PCR2 Custom ID: xxxxxx ^a	Blue	1 vial	350 µL ^b	
	IDTE pH 8.0	Blue	1 vial	2.0 mL	
BD Rhapsody Custom Panel 100–199 genes (Outside U.S.: Cat. No. 633778)	BD Rhapsody™ 10X PCR1 Custom ID: xxxxxx ^a	Blue	1 vial	1.4 mL ^b	-25°C to -15°C
	BD Rhapsody™ 10X PCR2 Custom ID: xxxxxx ^a	Blue	1 vial	350 µL ^b	
	IDTE pH 8.0	Blue	1 vial	2.0 mL	

Panel	Component	Cap color	Quantity	Volume	Storage
BD Rhapsody Custom Panel 200–299 genes (Outside U.S.: Cat. No. 633779)	BD Rhapsody™ 10X PCR1 Custom ID: xxxxxx ^a	Blue	1 vial	1.4 mL ^b	–25°C to –15°C
	BD Rhapsody™ 10X PCR2 Custom ID: xxxxxx ^a	Blue	1 vial	350 µL ^b	
	IDTE pH 8.0	Blue	1 vial	2.0 mL	
BD Rhapsody Custom Panel 300–399 genes (Outside U.S.: Cat. No. 633783)	BD Rhapsody™ 10X PCR1 Custom ID: xxxxxx ^a	Blue	1 vial	1.4 mL ^b	–25°C to –15°C
	BD Rhapsody™ 10X PCR2 Custom ID: xxxxxx ^a	Blue	1 vial	350 µL ^b	
	IDTE pH 8.0	Blue	1 vial	2.0 mL	
BD Rhapsody Custom Panel 400–500+ genes (Outside U.S.: Cat. No. 633784)	BD Rhapsody™ 10X PCR1 Custom ID: xxxxxx ^a	Blue	1 vial	1.4 mL ^b	–25°C to –15°C
	BD Rhapsody™ 10X PCR2 Custom ID: xxxxxx ^a	Blue	1 vial	350 µL ^b	
	IDTE pH 8.0	Blue	1 vial	2.0 mL	

- Each design has a unique identifier that is provided with your BD Rhapsody Custom Panel primer design.
- To prepare a 1X dilution for use in the assay, dilute 1 part PCR primer to 9 parts of IDTE buffer. Store the 1X dilution at –25°C to –15°C for ≤2 years.

**BD Rhapsody™
Panel Supplement
(Cat. No. 633742)**

Add up to 100 additional supplemental primers to the BD Rhapsody targeted (predesigned) or custom panels for a maximum of 500 primers that can be used in an experiment. To order a BD Rhapsody Panel Supplement, contact BD Biosciences at researchapplications@bd.com.

Up to two BD Rhapsody Panel Supplements can be added per reaction provided that the total number of primers added is ≤ 500 (predesigned or custom panel plus panel supplement) and the primers have been designed to be compatible.

Panel	Component	Cap color	Quantity	Volume	Storage
BD Rhapsody Panel Supplement (U.S. only: Cat. No. 633742)	BD Rhapsody™ 10X PCR1 Supp. ID: xxxxxx ^a	Blue	1 vial	120 μL^b	-25°C to -15°C
	BD Rhapsody™ 10X PCR2 Supp. ID: xxxxxx ^a	Blue	1 vial	30 μL^b	
	IDTE pH 8.0	Blue	1 vial	2.0 mL	
BD Rhapsody Panel Supplement 2–49 genes (Outside U.S.: Cat. No. 633770)	BD Rhapsody™ 10X PCR1 Supp. ID: xxxxxx ^a	Blue	1 vial	120 μL^b	-25°C to -15°C
	BD Rhapsody™ 10X PCR2 Supp. ID: xxxxxx ^a	Blue	1 vial	30 μL^b	
	IDTE pH 8.0	Blue	1 vial	2.0 mL	
BD Rhapsody Panel Supplement 50–100 genes (Outside U.S.: Cat. No. 633776)	BD Rhapsody™ 10X PCR1 Supp. ID: xxxxxx ^a	Blue	1 vial	120 μL^b	-25°C to -15°C
	BD Rhapsody™ 10X PCR2 Supp. ID: xxxxxx ^a	Blue	1 vial	30 μL^b	
	IDTE pH 8.0	Blue	1 vial	2.0 mL	

- Each design has a unique identifier that is provided with your BD Rhapsody Panel Supplement primer design.
- To prepare a 1X dilution for use in the assay, dilute 1 part PCR primer to 9 parts of IDTE buffer. Store the 1X dilution at -25°C to -15°C for ≤ 2 years.

Suggested kits from other vendors

Reagent	Supplier	Catalog no.
Agilent DNA High Sensitivity Kit	Agilent Technologies	5067-4626
High Sensitivity D1000 ScreenTape	Agilent Technologies	5067-5584
High Sensitivity D1000 Reagents	Agilent Technologies	5067-5585

Reagents

Required reagent for BD Rhapsody Cartridge workflow

Reagent	Supplier	Catalog no.
Absolute ethyl alcohol, molecular biology grade	Major supplier	—
Nuclease-free water	Major supplier	—

Required reagents for sample multiplexing and/or antibody- oligonucleotide labelling

Material	Supplier	Catalog no.
BD Pharmingen Stain Buffer (FBS)	BD Biosciences	554656
BD AbSeq Ab-Oligos	BD Biosciences	Various

Required reagents for cell preparation and staining

Reagent	Supplier	Catalog no.
Calcein AM cell-permeant dye ^a	Thermo Fisher Scientific	C1430
DRAQ7 TM ^a , 0.3 mM	BD Biosciences	564904
Dimethyl sulfoxide (DMSO)	Major supplier	—
70% ethyl alcohol or 70% isopropyl alcohol ^b	Major supplier	—

- Protect Calcein AM and DRAQ7 from light. Avoid multiple freeze-thaw cycles of Calcein AM. See manufacturer's storage recommendations.
- To clean the BD RhapsodyTM Express instrument and the BD RhapsodyTM Scanner, see the *BD RhapsodyTM Single-Cell Analysis System Installation and Maintenance Guide* (Doc ID: 43084). Instead of 70% alcohol, 10% (v/v) bleach can be used.

Required reagents for PCR product purification

Reagent	Supplier	Catalog no.
Agencourt® AMPure® XP magnetic beads	Beckman Coulter	A63880
Ethyl alcohol, Pure (200 proof, molecular biology grade)	Sigma-Aldrich	E7023-500ML

**Suggested reagent
for cell suspension
preparation**

Reagent	Supplier	Catalog no.
1X RBC Lysis Buffer	Thermo Fisher Scientific	00-4333-57
Phosphate-buffered saline (calcium- and magnesium-free)	Major supplier	—

**Suggested reagents
for sample
multiplexing and/or
antibody-
oligonucleotide
labelling**

Material	Supplier	Catalog no.
BD Pharmingen Human BD Fc Block ^a	BD Biosciences	564220

- a. For use with myeloid and B lymphocyte-containing samples.
-

Consumables

Required consumables

Consumable	Supplier	Catalog no.
Falcon® Tube with Cell Strainer Cap	Corning	352235
Falcon tubes, 5 mL Round Bottom Polystyrene Test Tube ^a	Corning	352054
Improved Neubauer Hemocytometer	INCYTO	DHC-N01-5
DNA LoBind Tubes, 1.5 mL ^b	Eppendorf	0030108051
DNA LoBind Tubes, 5 mL Note: These are the Bead Retrieval Tubes to be used with the BD Rhapsody Express instrument.	Eppendorf	0030108310
Low retention filtered pipette tips, 10 µL ^b	Major supplier	—
Low retention filtered pipette tips, 200 µL ^b	Major supplier	—
Low retention filtered pipette tips, 1,000 µL ^b	Major supplier	—

Consumable (continued)	Supplier	Catalog no.
Gilson™ PIPETMAN™ Tipack™ Filter Tips, 100–1,200 µL for BD Rhapsody™ P1200M pipette	Thermo Fisher Scientific	F171803G
Gilson PIPETMAN Tipack Racked Pipet Tips, 500–5,000 µL for BD Rhapsody™ P5000M pipette	Thermo Fisher Scientific	F161370G
Qubit™ Assay Tubes	Thermo Fisher Scientific	Q32856
Empty Latch Racks for 500 µL Tubes ^c	Thermo Fisher Scientific	4900 (4890 is also acceptable)
0.2 mL PCR 12-strip tubes ^a	Major supplier	—
10 mL sterile serological pipettes	Major supplier	—
Premoistened cleaning wipes with 70% ethyl alcohol or 70% isopropyl alcohol	Major supplier	—
Lint-free wipers	Major supplier	—

- a. Required for sample multiplexing and/or antibody-oligonucleotide labelling.
- b. Provide material in both pre- and post-amplification workspaces.
- c. Required for storing tubes of BD™ AbSeq Ab-Oligos and convenient pooling of reagents.

Suggested consumables

Consumable item	Supplier	Catalog no.
BD Vacutainer® CPT™ Mononuclear Cell Preparation Tube–Sodium Heparin ^a	BD Biosciences	362753

- a. For single cell preparation of peripheral blood mononuclear cells (PBMCs).
-

Equipment

Required equipment

Supply pre- and post-amplification workspaces with the required equipment. You might need two sets of some equipment.

Equipment	Box	Components
BD Rhapsody™ Single-Cell Analysis system	BD Rhapsody Scanner (Cat. No. 633701)	<ul style="list-style-type: none"> • BD Rhapsody Scanner • Power supply and cable • Hemocytometer Adapter (Cat. No. 633703) • BD Rhapsody P1200M pipette (Cat. No. 633704) • BD Rhapsody P5000M pipette (Cat. No. 633705)
	BD Rhapsody Express instrument (Cat. No. 633702)	<ul style="list-style-type: none"> • BD Rhapsody Express instrument

Equipment	Supplier	Catalog no.
6-Tube Magnetic Separation Rack for 1.5 mL tubes ^a	New England Biolabs	S1506S
Large magnetic separation stand	V&P Scientific, Inc.	VP 772FB-1
Clear acrylic cylinder adapter for 15 mL tube magnet ^b	V&P Scientific, Inc.	VP 772FB-1A
Low-profile magnetic separation stand for 0.2 mL, 8-strip tubes	V&P Scientific, Inc. OR	VP 772F4-1
	Clontech	635011
Thermomixer (37°C, 1,200 rpm): <ul style="list-style-type: none"> • ThermoMixer® C • SmartBlock™ Thermoblock 1.5 mL 	Eppendorf	<ul style="list-style-type: none"> • 5382000023 • 5360000038
Qubit™ 3.0 Fluorometer	Thermo Fisher Scientific	Q33216
Heat block capable of 80°C Suggested: VWR® Advanced Mini Dry Block Heater with Heated Lid	VWR	10153-348

Equipment (continued)	Supplier	Catalog no.
<ul style="list-style-type: none"> • 2100 Bioanalyzer OR • 4200 TapeStation Instrument 	Agilent Technologies	<ul style="list-style-type: none"> • G2940CA • G2991AA
Thermal cycler with heated lid	Major supplier	—
Water bath OR incubator at 37°C	Major supplier	—
Laminar flow hood	Major supplier	—
Digital timer ^a	Major supplier	—
Pipettes (P10, P20, P200, P1000) ^a	Major supplier	—
Multi-channel pipette, 2–20 µL OR 20–200 µL ^a	Major supplier	—
Microcentrifuge for 1.5–2.0 mL tubes ^a	Major supplier	—
Microcentrifuge for 0.2 mL tubes	Major supplier	—
Centrifuge and rotor with adapters for 5 mL Falcon tubes and 15 mL tubes	Major supplier	—
Vortexer ^a	Major supplier	—
Pipet-Aid	Major supplier	—

- a. Provide material in both pre- and post-amplification workspaces.
- b. Holds 5 mL LoBind Tube in magnet.

Suggested equipment

Equipment	Supplier	Catalog no.
Logitech® Wireless Combo (keyboard and mouse)	Major supplier	MK270
Barcode reader with keyboard wedge, USB 2.0 OR 3.0 compatible ^a	Major supplier	—
Phase-contrast microscope	Major supplier	—
8-Channel Screw Cap Tube Capper Note: Optional for antibody-oligonucleotide labelling	Thermo Fisher Scientific	4105MAT

- a. The external barcode reader is optional. The scanner has an internal barcode reader. To purchase an external barcode reader, BD Biosciences recommends the USB Automatic Barcode Scanner Scanning Barcode Reader with Hands-Free Adjustable Stand (Brainydeal) or equivalent.
-

Software

BD Rhapsody™ Analysis pipeline

The BD Rhapsody Analysis pipeline takes the FASTQ read files and reference files for gene alignment. The pipeline filters by read quality, annotates R1 and R2 reads, annotates molecules, determines putative cells, determines the sample of origin (sample multiplexing only), generates expression matrices, generates a metrics summary, and performs clustering analysis.

For installation, see the *BD Single Cell Genomics Analysis Setup User Guide* (Doc ID: 47383).

For detailed information on the BD Rhapsody Analysis pipeline, see the *BD Single Cell Genomics Bioinformatics Handbook* (Doc ID: 54169).

BD™ Data View

BD Data View is a software tool for visualization and exploratory analysis of output files generated following bioinformatics analysis. The software is included with the purchase of the BD Rhapsody™ Single-Cell Analysis system and is optional for use.

For installation, see the *BD Single Cell Genomics Analysis Setup User Guide* (Doc ID: 47383).

For detailed instructions on using BD Data View, see the *BD Single Cell Genomics Bioinformatics Handbook* (Doc ID: 54169).

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4

Preparing the BD Rhapsody™ Cartridge

- Priming the BD Rhapsody Cartridge (page 56)
- Treating the surface of the cartridge (page 63)

Priming the BD Rhapsody Cartridge

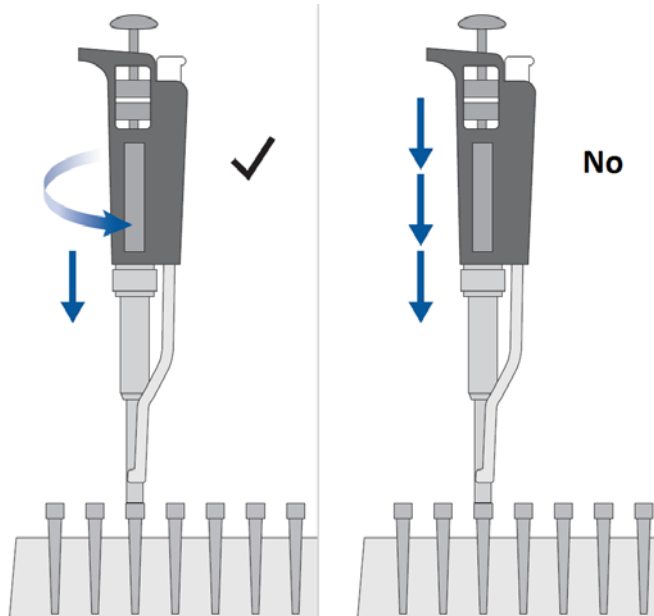
Before you begin After opening the cartridge packet, ensure that you prepare the cartridge in ≤ 1 day. If you leave the cartridge at 2°C to 8°C , equilibrate to room temperature (15°C to 25°C) for 5 minutes.

If cell preparation takes ≥ 4 hours, begin preparing cells before cartridge preparation. See the *Single Cell Analysis Workflow with BD Rhapsody™ Systems* (Doc ID: 220524) to find the appropriate protocol to follow.

- Equilibrate these reagents at room temperature (15°C to 25°C) for ≥ 30 minutes before use:
 - Cartridge Wash Buffer 1 (Cat. No. 650000060)
 - Cartridge Wash Buffer 2 (Cat. No. 650000061)
 - Place these reagents on ice:
 - Sample Buffer (Cat. No. 650000062)
 - 1 M DTT (Cat. No. 650000063)
 - Lysis Buffer (Cat. No. 650000064)
 - DRAQ7 (protected from light)
 - Once at room temperature (15°C to 25°C), resuspend Calcein AM (1 mg; Thermo Fisher Scientific Cat. No. C1430) in 503.0 μL of DMSO for a final stock concentration of 2 mM. Follow the manufacturer's storage recommendations, and protect it from light.
 - Review pipette settings and operation. See [Pipette Settings \(page 18\)](#).
 - For the use of two cartridges, see [Workflow with two BD Rhapsody™ Cartridges \(page 129\)](#).
-

Procedure

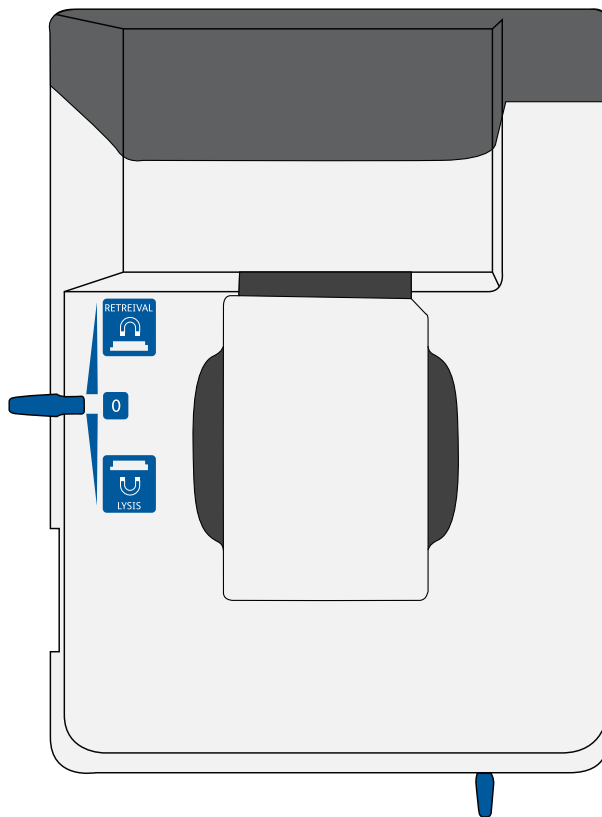
To ensure an air-tight seal with the BD Rhapsody™ P1200M or P5000M pipette, hold the pipette with one hand, and slightly twist the pipette to firmly seat a pipette tip on the pipette shaft:



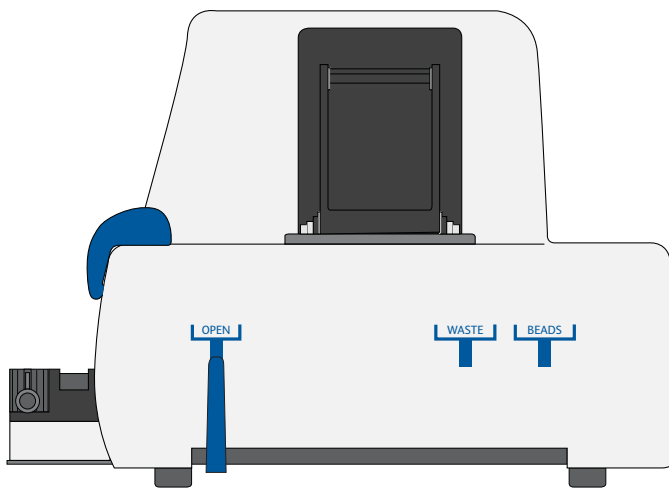
Avoid introducing bubbles while pipetting into the BD Rhapsody™ Cartridge.

Change pipette tips before every pipetting step.

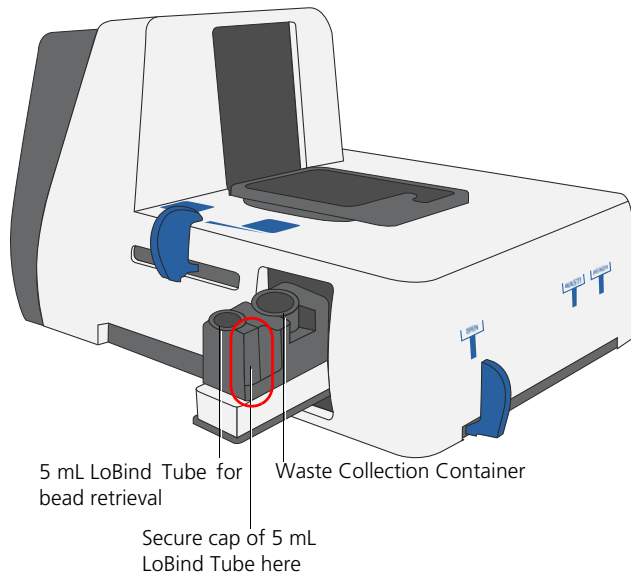
1. Move the left slider to the middle (0) position on the Express instrument. The Retrieval (top) magnet and Lysis (bottom) magnets are away from the cartridge tray:



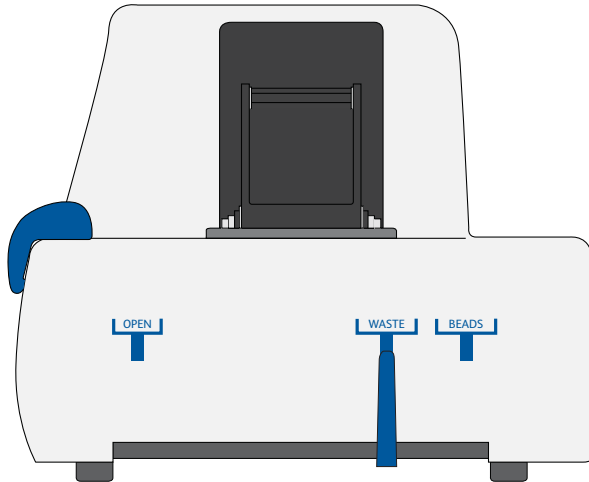
2. Move the front slider to **OPEN**:



3. Remove the cap of a Waste Collection Container (Cat. No. 650000090), and insert both the container and a new 5 mL LoBind Tube (Eppendorf Cat. No. 0030108310) for bead retrieval into the appropriate slots in the drawer. Secure the cap of the 5 mL LoBind Tube to the holder:

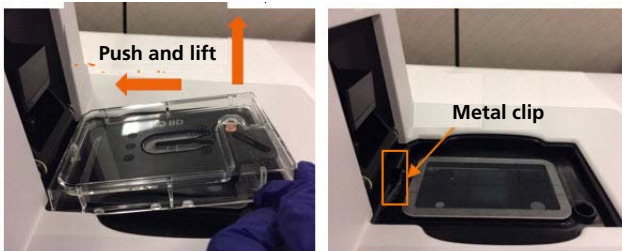


4. Move the front slider to **WASTE**:

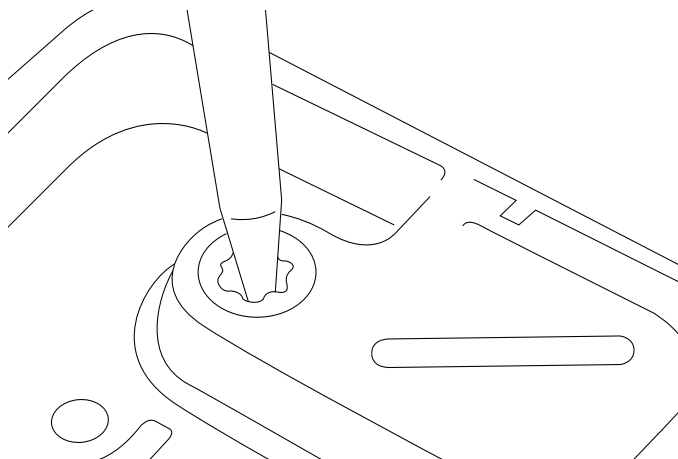


5. Push the cartridge into the far end of the tray to match cartridge and tray notches. Lay the cartridge flat, and release it. Ensure that the cartridge is flat in the tray and the barcode faces out.

Note: To remove the cartridge from the Express instrument, push in the cartridge, and lift it from the tray:



Before loading the reagent into the cartridge, align the pipette tip with the inlet hole of the gasket, and then press down on the P1200M pipette to seal the pipette tip against the gasket and avoid leaks:



Note: In **Prime/Treat** mode, press the button once to aspirate 700 μL , and press the button again to dispense 700 μL .

6. Load the cartridge with 700 μL of 100% (absolute) ethyl alcohol using the P1200M pipette in **Prime/Treat** mode.
 7. Load the cartridge with 700 μL of air using the P1200M pipette in **Prime/Treat** mode.
 8. Load the cartridge with 700 μL of Cartridge Wash Buffer 1 (Cat. No. 650000060) with the P1200M pipette in **Prime/Treat** mode.
 9. Leave the cartridge on the tray at room temperature (15°C to 25°C) for 1 minute.
-

Treating the surface of the cartridge

Procedure

1. Load the cartridge with 700 μ L of air using the P1200M pipette in **Prime/Treat** mode.
2. Load the cartridge with 700 μ L of Cartridge Wash Buffer 1 (Cat. No. 650000060) using the P1200M pipette in **Prime/Treat** mode.
3. Leave the cartridge on the tray at room temperature (15°C to 25°C) for 10 minutes.
4. Load the cartridge with 700 μ L of air using the P1200M pipette in **Prime/Treat** mode.
5. Load the cartridge with 700 μ L of Cartridge Wash Buffer 2 (Cat. No. 650000061) using the P1200M pipette in **Prime/Treat** mode.

Stopping point: The cartridge can be stored at room temperature (15°C to 25°C) for ≤ 4 hours. You can leave the cartridge on the tray. The performance of the cartridge has not been validated at room temperature (15°C to 25°C) storage for >4 hours.

6. Prepare a single cell suspension. See the *Single Cell Analysis Workflow with BD Rhapsody™ Systems* (Doc ID: 220524) to find the appropriate protocol to follow.
-

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5

Processing cells with the BD Rhapsody™ Single-Cell Analysis system

- Best practices for cell handling and cell counting (page 66)
- Staining cells with viability markers (page 67)
- Loading cells into the Hemocytometer Adapter (page 68)
- Counting cells with the BD Rhapsody Scanner (page 70)
- Preparing a cell suspension and loading cells in the BD Rhapsody™ Cartridge (page 77)
- Imaging cells in a cartridge (page 87)
- Preparing Cell Capture Beads (page 91)
- Loading Cell Capture Beads and imaging (page 92)
- Lysing cells and retrieving Cell Capture Beads (page 93)
- Performing reverse transcription on the Cell Capture Beads (page 99)
- Treating the sample with Exonuclease I (page 104)

Best practices for cell handling and cell counting

Cell handling

- Minimize cell handling to reduce cell loss and decline in cell viability.
 - Keep cells on ice when not handling them.
 - Optimize centrifugation conditions according to cell number and cell phenotype to see a cell pellet.
 - For high cell recovery, know the position of the cell pellet in the tube after centrifugation.
-

Cell counting

- Filter cells to remove clumps and debris to ensure accurate cell counting. Debris in suspensions of small cells can lead to overestimated cell counts. Visually check filtered cell suspensions for debris.
 - Avoid pipetting low volumes of cells (<2 μL).
 - Cell counting the BD Rhapsody™ Scanner is most accurate when the cell concentration is in the range of ~200–800 cells/ μL . If the cell concentration is >1,000 cells/ μL , dilute the cell suspension in cold Sample Buffer (Cat. No. 650000062) to ~200–800 cells/ μL .
 - Do not rely on FACS-based counts, because cell concentration might be overestimated by this method. Always re-count cells after FACS.
-

Staining cells with viability markers

- Before you begin**
- Prepare a single cell suspension. See the *Single Cell Analysis Workflow with BD Rhapsody™ Systems* (Doc ID: 220524) to find the appropriate protocol to follow.
If you are using biological samples that contain red blood cell contamination, red blood cell lysis is required. See the *Preparing Single Cell Suspensions Protocol* (Doc ID: 210964).
 - Thaw Calcein AM. Once at room temperature (15°C to 25°C), resuspend Calcein AM (1 mg; Thermo Fisher Scientific Cat. No. C1430) in 503.0 µL DMSO for a final stock concentration of 2 mM. Follow manufacturer's instructions, and protect from light.
-

Procedure

1. If cells are not resuspended in cold Sample Buffer (Cat. No. 650000062), centrifuge cell suspension at $400 \times g$ for 5 minutes, aspirate supernatant, and leave ~20 µL of residual supernatant. Add up to 620 µL total volume of cold Sample Buffer, and then proceed to step 2. If the total expected cell number is $\leq 30,000$ cells in 610 µL, proceed to step 2.

Performance might be impacted if samples are not in Sample Buffer. For rare samples that are not resuspended in Sample Buffer before cell loading, proceed at your own risk or contact tech support.

2. Add 3.1 µL of 2 mM Calcein AM (Thermo Fisher Scientific Cat. No. C1430) and 3.1 µL of 0.3 mM DRAQ7™ (BD Biosciences Cat. No. 564904) to 620 µL cell suspension (1:200 dilution) in cold Sample Buffer (Cat. No. 650000062).
3. Gently pipet-mix.
4. Incubate at 37°C in dark for 5 minutes.

5. Filter cells through Falcon® Tube with Cell Strainer Cap (Corning 352235).

For low abundance or low volume samples, filtering is optional at this step. BD Biosciences recommends filtering the final sample before loading cells into the cartridge.

6. Proceed immediately to [Loading cells into the Hemocytometer Adapter](#).

Loading cells into the Hemocytometer Adapter

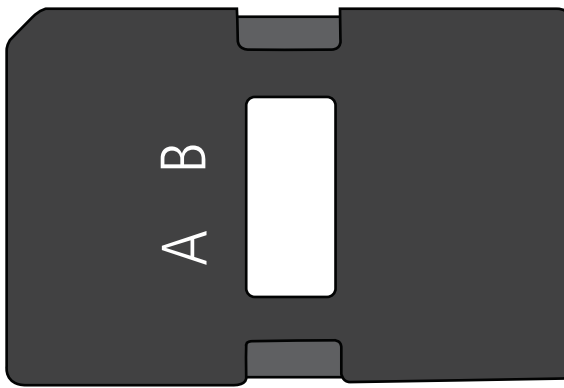
Before you begin Stain cells with viability markers. See [Staining cells with viability markers \(page 67\)](#).

Count cells immediately.

Loading cells Keep cells on ice, and protect them from light.

1. Gently mix cells well by pipette, and then gently pipet 10 µL of the cell suspension into one chamber of the INCYTO™ disposable hemocytometer (INCYTO Cat. No. DHC-N01-5).

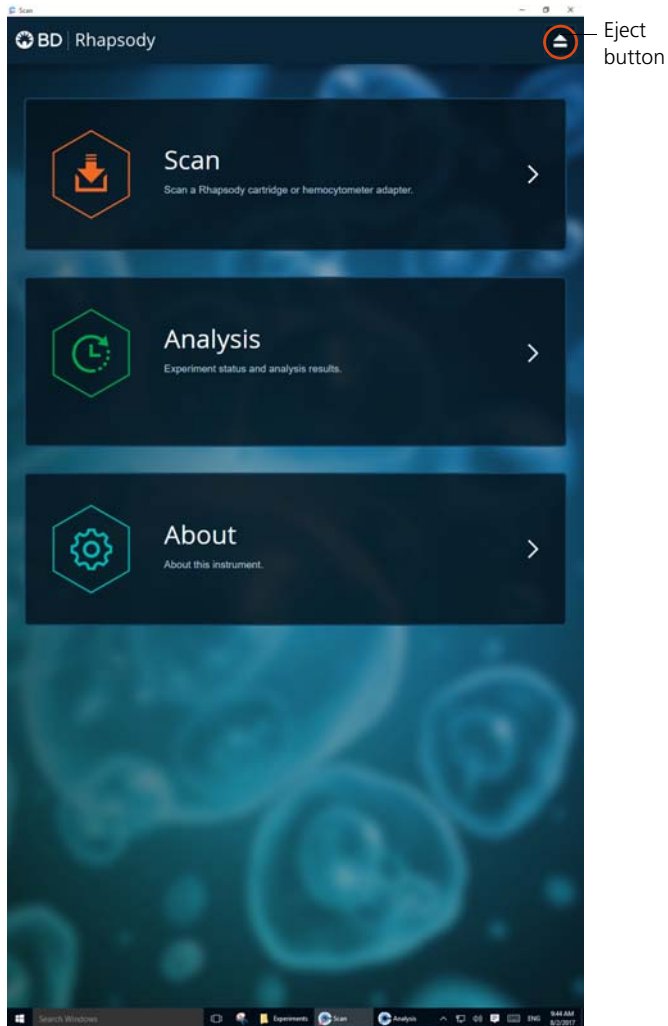
2. Insert the hemocytometer into the Hemocytometer Adapter (Cat. No. 633703) so that the A and B sides of the hemocytometer align with A and B on the Hemocytometer Adapter:



3. Count the cells in the BD Rhapsody Scanner ≤ 5 minutes after loading.
-

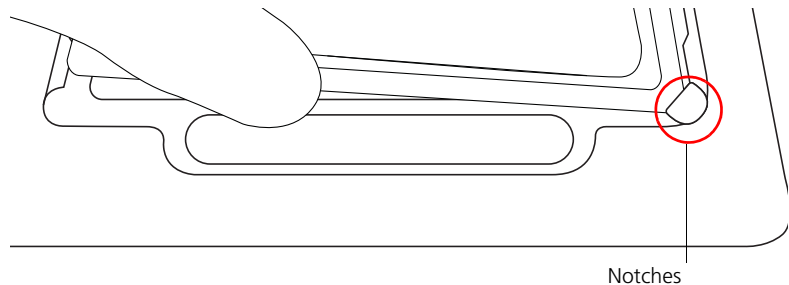
Counting cells with the BD Rhapsody Scanner

- Procedure**
1. Launch the scanner software so that the main menu displays:



Notes:

- To return to the main menu on any screen, tap the BD logo in the upper left.
 - For updates to the BD Rhapsody Scanner software, contact BD Biosciences technical support at researchapplications@bd.com.
 - To troubleshoot scanner software error messages, see [BD Rhapsody Scanner software messages \(page 114\)](#).
2. Tap **Scan**. The tray door of the scanner opens automatically, and the tray is ejected.
 3. Place the Hemocytometer Adapter on the scanner tray so that the notch of the adapter matches the corner notch, and the barcode faces toward the front of the instrument:

**Note:**

- The scanner displays an alert if the adapter is in the wrong orientation.
 - You can manually enter the barcode in the application.
4. Tap **Continue**. The tray retracts, the door closes, and the scanner displays the hemocytometer setup screen.

5. Select the protocol from the drop-down menu. Enter the experiment name, sample name, protocol, and user. (If an experiment name was entered previously, select it from the drop-down menu.)

Do not use commas in the experiment name, sample name, or user.

For example:

The screenshot displays the instrument's configuration interface. It features a dark blue header and footer. The main content area is white and contains several input fields:

- Experiment:** A dropdown menu with the value "20180131_SL_XH" selected.
- Sample:** A dropdown menu with the value "Jurkat" selected.
- Protocol:** A dropdown menu with the value "Hemocytometer" selected.
- User:** A text input field containing the value "SL".
- Cartridge:** A text input field containing the value "0107001033A".

Below the configuration fields, there are two sections:

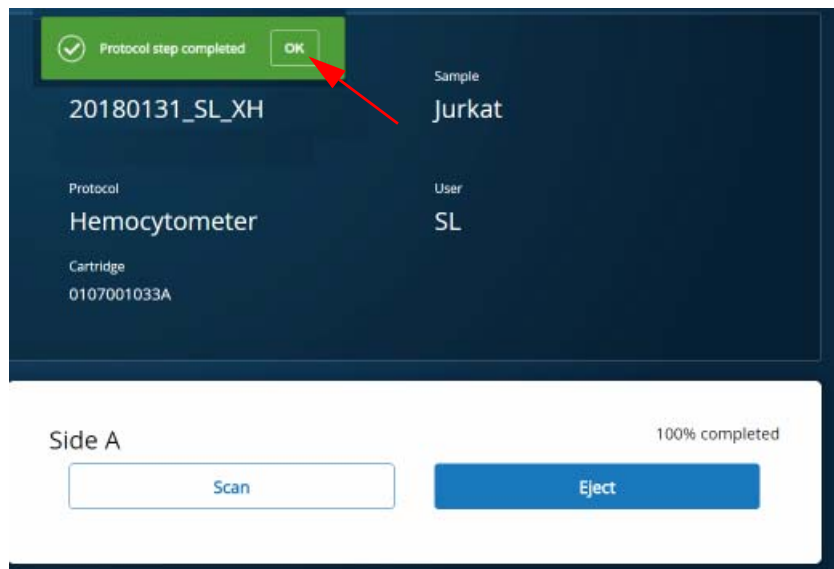
- Side A:** A white section with a blue button labeled "Start Side A Scan".
- Side B:** A dark blue section with the text "Side B" in white.

Note: Enter an experiment name that is appropriate for both the hemocytometer and cartridge scans. If necessary, tap the touch keyboard icon in the bottom right on the screen to enter information. There are no character or length limits.

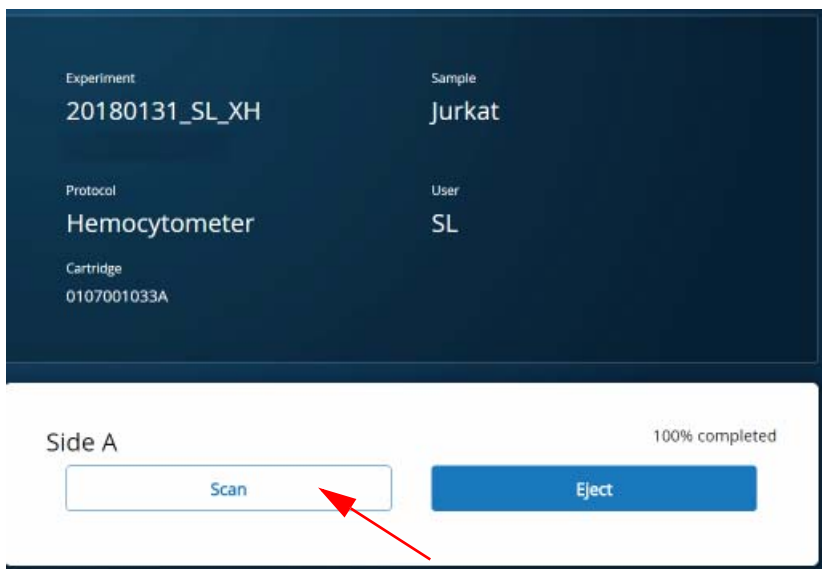
6. Tap **Side A** or **Side B**, depending on which side the cells were loaded, and then Select **Start Side A Scan** or **Start Side B Scan**, as required.

Note: To stop the scan, tap **Stop**, and then tap **Stop** again. To continue with the scan, tap **Continue**.

7. After the scan is complete, tap **OK**:



8. Scan the other side of the hemocytometer, or eject it:
 - Scan the other side: Tap **Scan**, enter a new sample name, and then repeat steps 6–7. The hemocytometer remains in the scanner and saves time waiting for the tray to eject. For example:



- Eject the hemocytometer: Tap **Eject**. The Hemocytometer Adapter is ejected from the scanner, and the remove cartridge screen displays.
Note: The analysis runs in the background. For example, the cell scan step for Jurkat and Ramos cells is complete. See step 11.
9. Remove the Hemocytometer Adapter from the tray, and then tap **Done** to retract the tray and return to the main menu.
 10. On the main menu, tap **Analysis**, and then tap the experiment. If no results are displayed, tap **Analyze**.

11. View the **total cell concentration** and **cell viability** on the results screen. For example, cell concentrations and viabilities are displayed for Jurkat and Ramos samples:

The image displays two screenshots of the BD Rhapsody Single-Cell Analysis system results screen. The top screenshot is for the 'Jurkat' sample, and the bottom screenshot is for the 'Ramos' sample. Both screenshots show a table with columns for Scan Date, Sample, Step, and Analysis Status. Below the table, there is an 'Analysis' section with two rows: 'Total cell concentration (live and dead)' and 'Cell viability'. The values for these metrics are highlighted with a red border in the original image.

Jurkat
Hemocytometer 0107001033A

Scan Date	Sample	Step	Analysis Status
2018-01-31 10:01:43.07	Jurkat	Side A	✓ Completed

Analysis

Total cell concentration (live and dead)	423.64
Cell viability	92.24

Ramos
Hemocytometer 0107001033A

Scan Date	Sample	Step	Analysis Status
2018-01-31 10:02:39.54	Ramos	Side B	✓ Completed

Analysis

Total cell concentration (live and dead)	375.99
Cell viability	92.53

Note: If no value could be calculated, then NaN (not a number) is displayed. See [Troubleshooting \(page 107\)](#).

12. Proceed as follows:
 - If the cell concentration is $\leq 1,000$ cells/ μ L, proceed to step [13](#).
 - If the cell concentration is $> 1,000$ cells/ μ L, dilute the cell suspension in cold Sample Buffer (Cat. No. 650000062) to ~ 200 – 800 cells/ μ L. Recount the cells in the hemocytometer following steps [1–11](#), and then proceed to step [13](#).
13. Tap **Prepare** at the top of the results screen to display the Samples Calculator screen.
14. Dispose of the hemocytometer according to local safety regulations. See [Preparing a cell suspension and loading cells in the BD Rhapsody™ Cartridge \(page 77\)](#).

Minimize the time between cell pooling and single cell capture.

Preparing a cell suspension and loading cells in the BD Rhapsody™ Cartridge

Best practices

- Always use low retention filtered pipette tips and LoBind Tubes.
 - Perform single cell capture and cDNA synthesis in a pre-amplification workspace.
 - Prepare cells as close to cell loading as possible. Keep other reagents, including Sample Buffer (Cat. No. 650000062) on ice unless instructed otherwise.
 - Change pipetting tips before every pipetting step.
-

Before you begin

- Prime and treat the BD Rhapsody Cartridge. See [Preparing the BD Rhapsody™ Cartridge \(page 55\)](#).
 - Thaw reagents (not enzymes) in the BD Rhapsody™ cDNA Kit at room temperature (15°C to 25°C), and then place them on ice. Keep enzymes at –25°C to –15°C.
 - Prepare a single cell suspension for cartridge loading.
 - Place these reagents on ice:
 - Sample Buffer (Cat. No. 650000062)
 - 1 M DTT (Cat. No. 650000063)
 - Lysis Buffer (Cat. No. 650000064)
 - Cell Capture Beads (Cat. No. 650000089)
-

Loading cells into the cartridge

To ensure an air-tight seal with the BD Rhapsody™ P1200M and P5000M pipettes, hold the pipette with one hand, and slightly twist the pipette to firmly seat a pipette tip on the pipette shaft.

1. Determine the desired number of cells to capture in the BD Rhapsody Cartridge. The following table lists the estimated multiplet rate based on the number of captured cells on retrieved Cell Capture Beads:

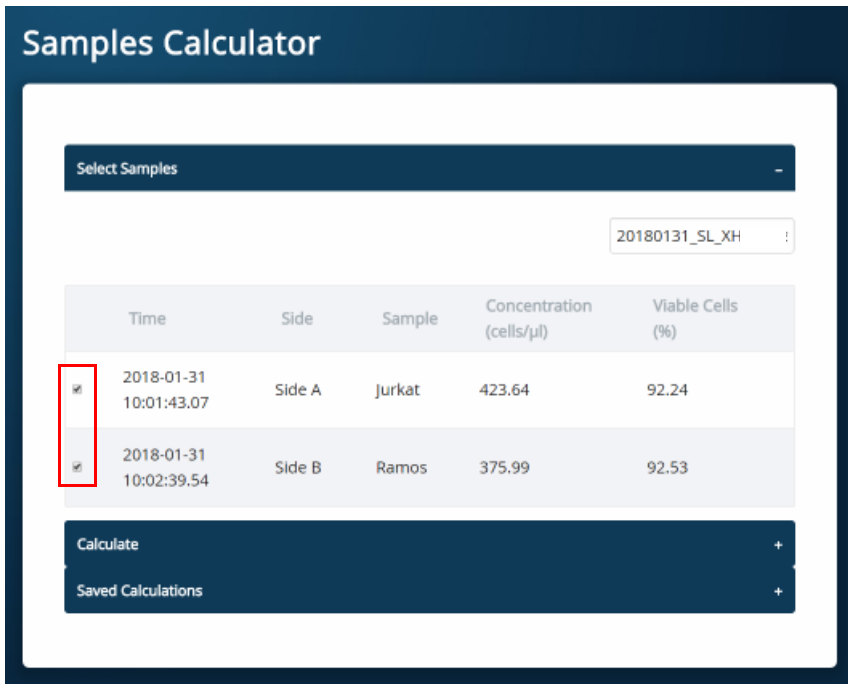
Estimated multiplet rate based on the number of captured cells on retrieved Cell Capture Beads

Number of captured cells on retrieved Cell Capture Beads (target)^{a,b}	Estimated multiplet rate (%)
100	0.0
500	0.1
1,000	0.2
2,000	0.5
3,000	0.7
4,000	1.0
5,000	1.2
6,000	1.4
7,000	1.7
8,000	1.9
9,000	2.1
10,000	2.4
11,000	2.6
12,000	2.8
13,000	3.1
14,000	3.3
15,000	3.5
16,000	3.8
17,000	4.0

Number of captured cells on retrieved Cell Capture Beads (target) ^{a,b} (continued)	Estimated multiplet rate (%)
18,000	4.2
19,000	4.5
20,000	4.7

- a. The number of cells sequenced might be less than the number of cells captured due to bead loss during handling, panel choice, and sample composition. The validated range of cells sequenced is 100–10,000 cells.
 - b. This sample calculator gives loading calculations based on *total cell count*, which does not consider cell viability. The number of viable cells captured in the cartridge might be less than the targeted number of captured cells if the viability of the sample is <100%.
2. According to the number of captured cells chosen in step 1, use the Samples Calculator on the scanner to obtain the volume(s) of stock cell and volume of cold Sample Buffer (Cat. No. 650000062) to prepare a cell suspension of 650 µL for loading into a BD Rhapsody Cartridge.
 3. In the BD Rhapsody Scanner software, navigate to the **Analysis** screen, and proceed to the next step. If you are on the results screen that displays the viability and concentration of cells, tap **Prepare** at the top of the screen, and skip to step c.
 - a. On the Analysis screen, tap the experiment.
 - b. On the results screen, tap **Prepare**.
 - c. On the Samples Calculator screen, select the experiment from the drop-down menu.
 - d. You can search an experiment on the current screen by entering keywords in the search box.

- e. If necessary, tap the check boxes to de-select samples. For example, two samples are selected:



- f. Tap **Calculate**.
- g. Select the cartridge type from the drop-down menu. The cartridge type is the first four digits on the barcode label of the cartridge.

- h. Enter the **Desired total volume** and the **Desired number of captured cells**. For example, a desired total volume of 650 μL of a mixture of two samples and 10,000 cells captured are entered:

Samples Calculator

Select Samples +

Calculate -

Cartridge type
0109 575 μl

Desired total volume - 650 + μl

Desired number of captured cells - 10000 + cells

Sample	Time	Concentration (cells/ μl)	Viable Cells (%)	Relative Amounts
--------	------	---------------------------------------	------------------	------------------

- i. If necessary, enter the relative amount of each sample below the desired number of captured cells. For example, equal amounts of two samples are entered:

Desired number of captured cells cells

Sample	Time	Concentration (cells/ μ l)	Viable Cells (%)	Relative Amounts
Jurkat	2018-01-31 10:01:43.07	423.64	92.24	<input type="button" value="-"/> <input type="text" value="1"/> <input type="button" value="+"/>
Ramos	2018-01-31 10:02:39.54	375.99	92.53	<input type="button" value="-"/> <input type="text" value="1"/> <input type="button" value="+"/>

Results

- j. Obtain the calculated stock cell and buffer volumes to prepare the cell suspension for loading into the cartridge.

For example, two stock volumes and the buffer volume are calculated:

Results	
Sample	Stock Volume
Jurkat	15.3 µl
Ramos	17.2 µl
Buffer volume	617.5 µl
Loading cell concentration	19.9 cells/µl
Estimated cell doublet rate	2.4%

Save

- k. (Optional) Tap **Save**. You can view the calculation by clicking the **Rhapsody Data** shortcut on the scanner screen and navigating to the appropriate folder.

1. (Optional) Click **Saved Calculations**. You can click saved calculations at any time. For example, the saved calculations from one experiment with two samples are displayed:

Samples Calculator

Select Samples +
Calculate +
Saved Calculations -

Cartridge type 0109	575 μ l	Buffer volume	617.5 μ l
Desired total volume	650 μ l	Loading cell concentration	19.9 cells/ μ l
Desired number of captured cells	10000 cells	Estimated cell doublet rate	2.4 %

Sample	Operator	Scan Time	Concentration (cells/ μ l)	Viable Cells (%)	Relative Amounts	Stock Volume
Jurkat		2018-01-31 10:01:43 .07	423.6	92.24	1	15.3
Ramos		2018-01-31 10:02:39 .54	376	92.53	1	17.2

4. Prepare the cell suspension:

Stock cell volume or cell number	Cell suspension preparation
Calculated Stock Volume is <610 μL	In a new 1.5 mL LoBind Tube, prepare the cell suspension in cold Sample Buffer (Cat. No. 650000062) according to the volumes obtained in step 3.
<ul style="list-style-type: none"> • Calculated Stock Volume $\geq 610 \mu\text{L}$, or • Buffer volume is negative, or • Cell concentration too low warning is displayed 	Use 610 μL of the cold cell suspension for cartridge cell loading. ^a

- a. If the ratio of cells and targeted number of cells cannot be achieved, mix cells to prepare the maximum input of cells in a total volume of 610 μL . For example, if the Samples Calculator specifies mixing 700 μL of Jurkat cells and 30 μL of Ramos cells, mix 580 μL of Jurkat cells with 30 μL of Ramos cells.

Ensure the stock solution is well suspended by gently pipetting cells up and down before transferring the appropriate volume for dilution.

5. If the samples were not filtered before counting cells, pass the final dilution of the prepared single sample or pooled sample through a Falcon® Tube with Cell Strainer Cap (Thermo Fisher Scientific Cat. No. 352235) before loading the sample into the cartridge.
6. Load the cartridge on the tray with 700 μL of air using the BD Rhapsody P1200M pipette in **Prime/Treat** mode.
7. Change the mode of the BD Rhapsody P1200M pipette to **Cell Load**.

8. With a manual pipette, gently pipet the cell suspension up and down to mix.
9. On the BD Rhapsody P1200M pipette, press the pipette button once to aspirate 40 μL of air, immerse the pipette tip in cell suspension, and then press the button again to aspirate 575 μL of cold cell suspension.
10. Insert the tip of the pipette perpendicular to the port, seal the pipette tip against the gasket, and then press the button a third time to dispense 615 μL of air and cells.

Air bubbles that might appear at the inlet or outlet of the cartridge do not affect cartridge performance.

11. Incubate the cells by one of these methods:
 - On the BD Rhapsody™ Express instrument: Leave the cartridge with loaded cells on the tray at room temperature (15°C to 25°C) for 15 minutes. During incubation on the laboratory bench, prepare the Cell Capture Beads (Cat. No. 650000089). See [Preparing Cell Capture Beads \(page 91\)](#). After preparing the Cell Capture Beads, proceed to [Imaging cells in a cartridge](#), or
 - In the scanner: Proceed immediately to [Imaging cells in a cartridge \(page 87\)](#) to install the cartridge and enter a 15 minute run delay.
-

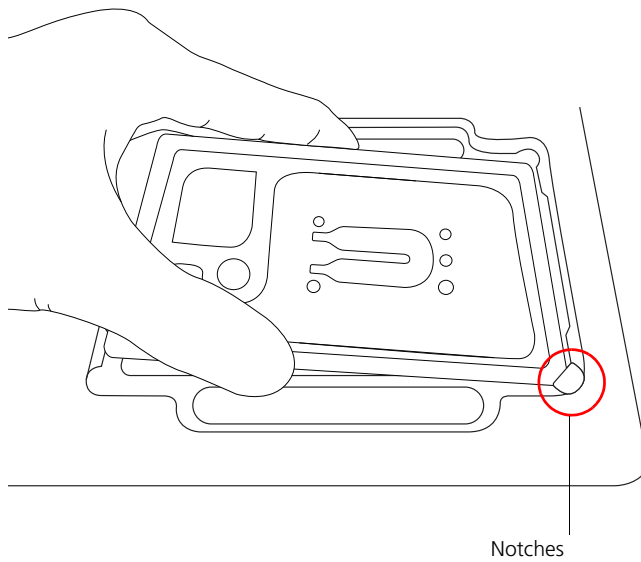
Imaging cells in a cartridge

Procedure

1. Navigate to the **Scan** application. The tray door of the scanner opens automatically, and the tray is ejected.

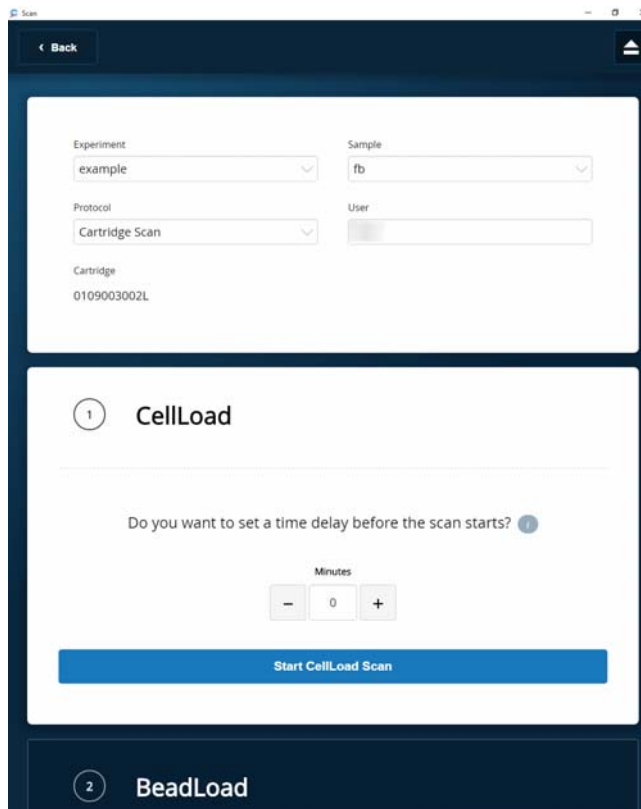
Note: If the tray is not ejected, tap the eject button in the upper right of the cartridge insertion screen.

2. Push the cartridge into the far end of the Express instrument tray to match the cartridge and tray notches. Lay the cartridge flat, and release it. Ensure that the cartridge is flat in the tray and the barcode faces out.
3. If necessary, wipe condensation from the top surface of the cartridge with a lint-free wiper to ensure optimal scanning.
4. Place the cartridge on the scanner tray so that the cartridge and tray notches match and the barcode faces toward the front of the instrument:



Notes:

- The scanner displays an alert if the cartridge is in the wrong orientation.
 - You can manually enter the barcode in the application.
5. Tap **Continue**. The tray retracts, the door closes, and the scanner displays the experiment setup screen.
 6. Select from the drop-down menu or enter the experiment name, sample name, protocol, and user.
 7. Tap **Cell Load**:



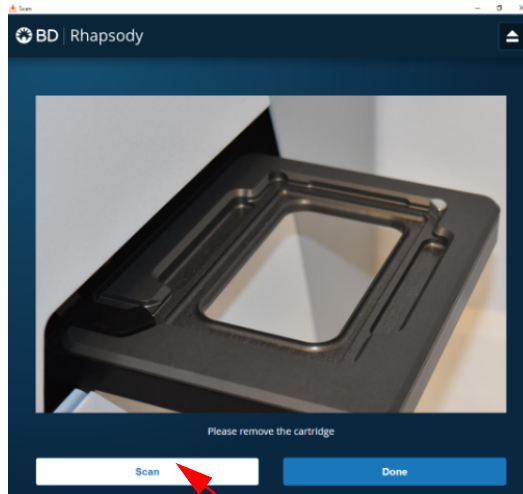
8. Proceed according to incubation location:
 - Express instrument (15 minute incubation completed): Proceed to step 9.
 - Incubation in scanner: Enter a 15 minute time delay to delay the start of the scan, and incubate the cartridge in the scanner by proceeding to step 9. During incubation in the scanner, prepare the Cell Capture Beads (Cat. No. 650000089). See [Preparing Cell Capture Beads \(page 91\)](#). After preparing the Cell Capture Beads, proceed to step 10.
9. Tap **Start Cell Load Scan**. The tray retracts into the scanner, and the door closes. If entered, the 15 minute countdown time displays in the Cell Load window. The scan proceeds after the countdown completes. The scan takes an additional ~4 minutes to complete.

Note: To stop the scan, tap **Stop**, and then tap **Stop** again. You can then scan the cartridge with a different step or rerun the scan. To continue with the current scan, tap **Continue**.
10. After the scan is complete, tap **OK**, and then **Eject**. The cartridge is ejected from the scanner, and the remove cartridge screen is displayed.

Note: The analysis runs in the background.

11. Scan another cartridge, or retract the tray:

- Scan another cartridge: Tap **Scan**, and proceed to step 4 to place a new cartridge on the tray for scanning:



- Retract the tray: Tap **Done**

12. To confirm that the analysis is running, navigate to the **Analysis** screen, and then tap the experiment to view the ongoing analysis on the results screen. Upon completion, the results are listed:

Ctrl-85pct

Cartridge 0109033258A

Scan Date	Sample	Step	Analysis Status
2018-01-31 10:26:00.49	Ctrl-85pct	Cell Load	✓ Completed

Note: If the analysis is not running, tap **Analyze**. To re-analyze a scan, tap the row for the scan, and then tap **Re-Analyze**.

Preparing Cell Capture Beads

- Before you begin**
- Prepare the pre-amplification workspace for preparation of the Cell Capture Beads for the BD Rhapsody Cartridge.
 - Keep the Cell Capture Beads on ice before use.
 - For maximum recovery, do not vortex samples containing Cell Capture Beads.
 - Gently mix suspensions with Cell Capture Beads by pipette only.
-

Preparing Cell Capture Beads

Use low retention pipette tips and LoBind Tubes when handling Cell Capture Beads.

1. Place the tube with Cell Capture Beads (Cat. No. 650000089) (beads) on the 1.5 mL tube magnet for 1 minute.
 2. Carefully remove and appropriately discard the storage buffer without disturbing the beads and while leaving the tube on the magnet.
 3. Remove the tube from the magnet, and then pipet 750 μ L of cold Sample Buffer (Cat. No. 650000062) into the tube of beads.
 4. Pipet the bead suspension up and down to mix.
 5. Keep the beads on ice.
 6. After the Cell Load scan, and after you have confirmed that the analysis is running, proceed to [Loading Cell Capture Beads and imaging \(page 92\)](#).
-

Loading Cell Capture Beads and imaging

1. Return the cartridge to the tray of the BD Rhapsody Express instrument.
2. Change the mode of the BD Rhapsody P1200M pipette to **Prime/Treat**.
3. Load the cartridge with 700 μL of air using the BD Rhapsody P1200M pipette in **Prime/Treat** mode.
4. Change the mode of the BD Rhapsody P1200M pipette to **Bead Load**.
5. Use a P1000 standard pipette to gently pipet the Cell Capture Beads in cold Sample Buffer (Cat. No. 650000062) up and down to mix, and, using the BD Rhapsody P1200M pipette in **Bead Load** mode, immediately load the cartridge with 630 μL of beads.
6. Let the beads settle in the cartridge on the tray at room temperature (15°C to 25°C) for 3 minutes.
7. Image the cartridge with the scanner (**Bead Load** step). For detailed instructions, see [Imaging cells in a cartridge \(page 87\)](#).
8. Return the cartridge to the tray of the BD Rhapsody Express instrument.
9. Change the mode of the BD Rhapsody P1200M pipette to **Wash**.

Note: In **Wash** mode, press the button once to aspirate 720 μL of air or reagent. After aspiration, insert the tip into the cartridge, and then press the button once to dispense 700 μL of air or liquid. After removing the pipette tip from the cartridge inlet, press the button once to dispense the remaining 20 μL of air or liquid before ejecting the pipette tip.

10. Load the cartridge with 700 μL of air using the BD Rhapsody P1200M pipette in **Wash** mode.
 11. Load the cartridge with 700 μL of cold Sample Buffer (Cat. No. 650000062) using the BD Rhapsody P1200M pipette in **Wash** mode.
 12. Repeat steps 10–11 once for a total of two washes.
 13. Image the cartridge with the scanner (**Bead Wash** step). For detailed instructions, see [Imaging cells in a cartridge \(page 87\)](#).
-

Lysing cells and retrieving Cell Capture Beads

Lysing the cells

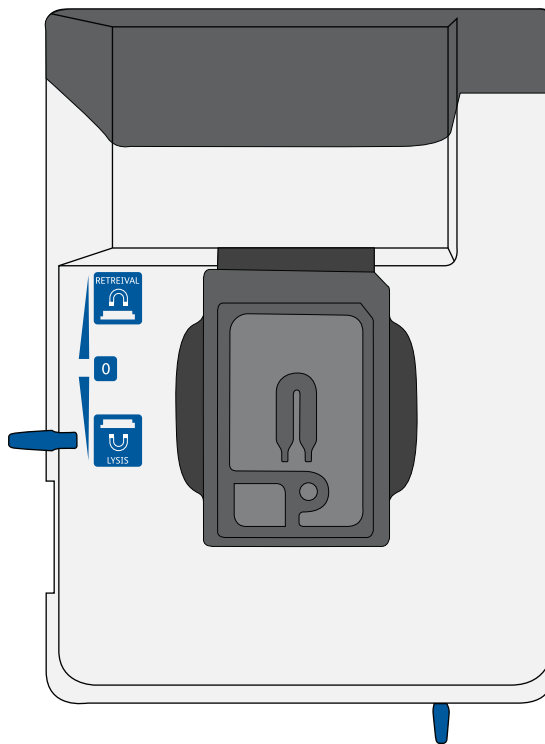
Avoid bubbles.

1. Add 75.0 μL of 1 M DTT (Cat. No. 650000063) to one bottle of 15 mL Lysis Buffer (Cat. No. 650000064), and then check the **Add DTT** box on the Lysis Buffer label.

Use the Lysis Buffer with DTT \leq 24 hours, and then discard.

2. Briefly vortex the lysis mix, and place it on ice.
3. Return the cartridge to the tray of the BD Rhapsody Express instrument.

4. Move the left slider to **LYSIS**. The (bottom) magnet is in the up position and is in contact with the cartridge:

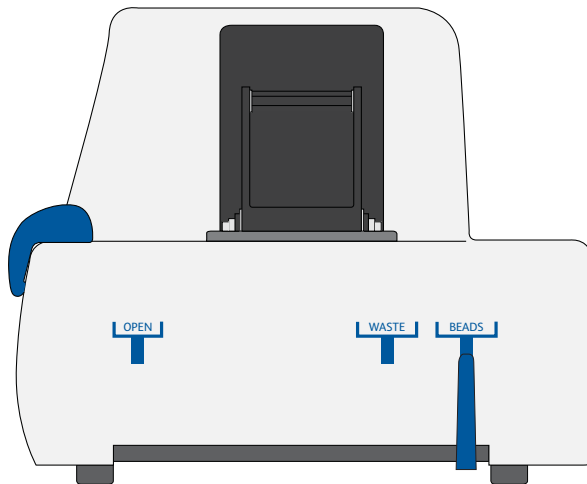


5. Change the mode on the BD Rhapsody P1200M pipette to **Lysis**.
6. Load the cartridge with 550 μL of Lysis Buffer with DTT using the BD Rhapsody P1200M pipette in **Lysis** mode.
7. Leave the cartridge at room temperature (15°C to 25°C) on the tray for 2 minutes.

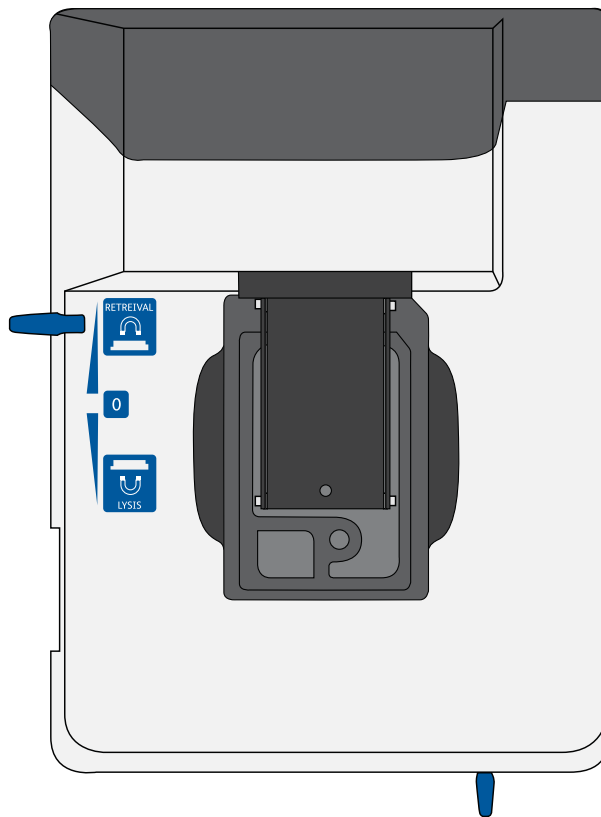
Maintain recommended lysis time for best performance.

Retrieving the Cell Capture Beads from the cartridge

1. Ensure that a 5 mL LoBind Tube (Eppendorf Cat. No. 0030108310) was inserted into the drawer for bead retrieval.
2. Confirm that the mode on the BD Rhapsody P5000M pipette is **Retrieval**. The pipette is locked into this single mode.
3. Move the front slider to **BEADS**:



4. Move the left slider to **RETRIEVAL**. The (top) magnet is in the down position and is in contact with the cartridge:



5. Leave the Retrieval magnet in the down position for 30 seconds.
6. Use the BD Rhapsody P5000M pipette to aspirate 5,000 μL of Lysis Buffer with DTT.
7. Press down firmly on the BD Rhapsody P5000M pipette to seal the pipette tip against the gasket of the cartridge to avoid leaks.


8. Move the left slider to the middle (0) position, and *immediately* load the cartridge with 4,950 µL of Lysis Buffer with DTT using the BD Rhapsody P5000M pipette. The Retrieval (top) magnet is in its full up position and is away from the cartridge.

The Cell Capture Beads (beads) are collected in the 5 mL LoBind Tube.

9. Remove the pipette tip from the inlet gasket of the cartridge before pressing the dial button once to purge the tip. Discard the pipette tip.
10. Move the front slider to **OPEN**, and then remove and cap the 5 mL LoBind Tube.
11. Uncap the 5 mL LoBind Tube, and place it on the large magnetic separation stand fitted with the 15 mL tube adapter for 1 minute.

During the 1 minute incubation, proceed to imaging the cartridge in step 12.

12. Image the cartridge with the scanner (**Retrieval** step). For detailed instructions, see [Imaging cells in a cartridge \(page 87\)](#).
13. Proceed immediately to [Performing reverse transcription on the Cell Capture Beads \(page 99\)](#) to process the beads and begin reverse transcription.
14. Appropriately dispose of the BD Rhapsody Cartridge according to biosafety level (BSL):

	Biological hazard.
---	---------------------------

- BSL-1. Discard the cartridge in a recycle container.
- BSL-2. Discard the cartridge in a biosafety waste container.

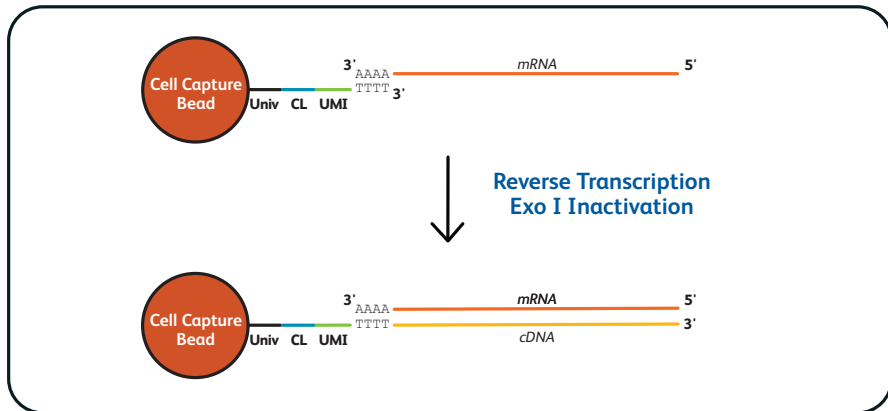
Dispose of waste using proper precautions and in accordance with local regulations. For more information, see [Waste \(page 140\)](#).

15. Appropriately dispose of the waste in the Waste Collection Container.
 16. Appropriately dispose of the Lysis Buffer with DTT.
 17. Wipe the Express instrument with 10% bleach or 70% ethyl alcohol. See the *BD Rhapsody™ Single-Cell Analysis System Installation and Maintenance Guide* (Doc ID 43084).
-

Performing reverse transcription on the Cell Capture Beads

Introduction

Prepare the reverse transcription mix, wash the Cell Capture Beads, and then perform reverse transcription on the beads with captured polyadenylated targets.



Univ: universal oligo; CL: cell label; UMI: Unique Molecular Identifier.

Best practices

- Prepare the cDNA mix in the pre-amplification workspace.
- Start reverse transcription ≤ 30 minutes after washing retrieved beads with Bead Wash Buffer.

Before you begin

- Obtain the 5 mL LoBind Tube of retrieved beads. See [Retrieving the Cell Capture Beads from the cartridge \(page 95\)](#).
- Ensure that the SmartBlock™ Thermoblock 1.5 mL or equivalent is installed on the thermomixer and is set to 37°C and 20 minutes.

Washing the Cell Capture Beads

Keep the Cell Capture Beads cold during washes.

Use low retention tips to handle Cell Capture Beads.

1. After the 1 minute incubation on the large magnet [see [Retrieving the Cell Capture Beads from the cartridge \(page 95\)](#)] and while leaving the 5 mL LoBind Tube on the large magnet, use a pipette to carefully remove all but ~1 mL of supernatant without disturbing the beads.
2. Remove the tube from the large magnet, resuspend the ~1 mL beads by gently pipetting the suspension up and down, and then transfer the bead suspension to a new 1.5 mL LoBind Tube.
3. If beads remain in the 5 mL LoBind Tube, pipet an additional 0.5 mL of Lysis Buffer with DTT into the tube, rinse the 5 mL tube, and transfer the suspension to the 1.5 mL LoBind Tube of beads.
4. Place the tube on the 1.5 mL tube magnet for ≤ 2 minutes, and then carefully remove and appropriately discard the supernatant without disturbing the beads and while leaving the tube on the magnet.

Avoid leaving Lysis Buffer or bubbles in the tube. Lysis Buffer might cause the reverse transcription reaction to fail.

5. Remove the 1.5 mL LoBind Tube from the magnet, and then pipet 1.0 mL of cold Bead Wash Buffer (Cat. No. 650000065) into the tube. Gently mix the suspension by pipette only. Do not vortex.

6. Place the tube on the 1.5 mL tube magnet for ≤ 2 minutes, and then carefully remove and appropriately discard the supernatant without disturbing the beads and while leaving the tube on the magnet.
7. Remove the 1.5 mL LoBind Tube from the magnet, and then pipet 1.0 mL of cold Bead Wash Buffer (Cat. No. 650000065) into the tube. Gently mix the suspension by pipette only, and place the tube on ice. Do not vortex.

Start reverse transcription ≤ 30 minutes after washing retrieved Cell Capture Beads with Bead Wash Buffer.

Performing reverse transcription

When working with Cell Capture Beads (beads), use only low retention tips and LoBind Tubes.

Limit preparation of mixes to $\leq 20\%$ overage.

Prepare the cDNA mix on ice.

1. Ensure that the SmartBlock Thermoblock for ThermoMixer® C is at 37°C.
2. In the pre-amplification workspace, into a new 1.5 mL LoBind Tube that is on ice, pipet the components in the following order to prepare the cDNA mix:

cDNA mix

Component	1 library (µL)	1 library + 20% overage (µL)
RT Buffer (Cat. No. 650000067)	40.0	48.0
dNTP (Cat. No. 650000077)	20.0	24.0
RT 0.1 M DTT (Cat. No. 650000068)	10.0	12.0
Bead RT/PCR Enhancer (Cat. No. 91-1082)	12.0	14.4
RNase Inhibitor (Cat. No. 650000078)	10.0	12.0
Reverse Transcriptase (Cat. No. 650000069)	10.0	12.0
Nuclease-Free Water (Cat. No. 650000076)	98.0	117.6
Total	200.0	240.0

3. Gently vortex and centrifuge the mix, and then place it back on ice.

4. Place the tube of washed beads [see [Washing the Cell Capture Beads \(page 100\)](#)] on the 1.5 mL tube magnet for ≤ 2 minutes, and then carefully remove and appropriately discard the supernatant without disturbing the beads and while leaving the tube on the magnet.
5. Use a low retention tip to pipet 200 μL of the cDNA mix to resuspend the beads. Gently mix the suspension by pipette only. Do not vortex.

Prepared cDNA mix with beads should be kept on ice until the suspension is transferred in the next step.

6. Transfer the bead suspension to a new 1.5 mL LoBind Tube.
7. Incubate the suspension on the thermomixer at 1,200 rpm and 37°C for 20 minutes.

Shaking is critical for this incubation.

During reverse transcription incubation, view the image analysis to determine if the analysis metrics have passed. See [Reviewing the analysis metrics \(page 124\)](#).

8. After incubation, place the tube on ice.
-

Treating the sample with Exonuclease I

- Before you begin**
- Ensure that the SmartBlock Thermoblock 1.5 mL or equivalent is installed on the thermomixer and is set to 37°C and 30 minutes.
 - Set a second thermomixer or heat block to 80°C.
-

Preparing the Exonuclease I mix When working with Cell Capture Beads, use only low retention tips and LoBind Tubes.

Limit preparation of mixes to $\leq 20\%$ overage.

Prepare the Exonuclease I mix on ice.

1. In the pre-amplification workspace, prepare the Exonuclease I mix in a new 1.5 mL LoBind Tube that is on ice by adding the components in the following order:

Exonuclease I mix

Component	1 library (μL)	1 library + 20% overage (μL)
10X Exonuclease I Buffer (Cat. No. 650000071)	20.0	24.0
Exonuclease I (Cat. No. 650000072)	10.0	12.0
Nuclease-Free Water (Cat. No. 650000076)	170.0	204.0
Total	200.0	240.0

2. Gently vortex and centrifuge the mix, and then place it back on ice.
-

**Treating the Cell
Capture Beads with
Exonuclease I**

1. Place the tube of beads with cDNA mix on the 1.5 mL tube magnet for ≤ 2 minutes, and then carefully remove and appropriately discard the supernatant without disturbing the beads and while leaving the tube on the magnet.
2. Remove the tube from the magnet, and then use a low retention tip to pipet 200 μL of Exonuclease I mix into the tube, Gently resuspend the beads by pipette only. Do not vortex.
3. Incubate the suspension on the thermomixer at 1,200 rpm and 37°C for 30 minutes.
4. If the thermomixer or heat block needs to preheat to a different temperature (80°C thermomixer or heat block), place the samples on ice until that temperature is reached.
5. Immediately proceed to [Inactivating Exonuclease I](#).

**Inactivating
Exonuclease I**

1. Transfer the bead suspension with Exonuclease I to the thermomixer (no shaking) in the pre-amplification workspace at 80°C for 20 minutes, or place the bead suspension in a heat block at 80°C for 20 minutes.
2. Place the bead suspension on ice for ~1 minute.
3. Place the tube on the 1.5 mL tube magnet until the solution is clear (≤ 1 minute).
4. Carefully remove and appropriately discard the supernatant without disturbing the beads and while leaving the tube on the magnet.

5. Remove the tube from the magnet, and with a low retention tip, pipet 200 μL of cold Bead Resuspension Buffer (Cat. No. 650000066) to gently resuspend the beads. Do not vortex.

Stopping point: The Exonuclease I-treated beads can be stored at 2°C to 8°C for ≤ 3 months.

6. Proceed to library preparation. See the *Single Cell Analysis Workflow with BD Rhapsody™ Systems* (Doc ID: 220524) to find the appropriate protocol to follow.
-

6

Troubleshooting

- Scanning troubleshooting (page 108)
- Cartridge loading troubleshooting (page 110)
- BD Rhapsody Scanner software messages (page 114)

Scanning troubleshooting

Introduction This topic describes possible problems and recommended solutions for scanning issues.

Incorrect alignment

Possible causes	Recommended solutions
Cartridge or hemocytometer not properly inserted on BD Rhapsody™ Express instrument tray	Ensure that the notched corner of the cartridge or hemocytometer is aligned with the notch of the tray, and the barcode is facing toward the front of the instrument.

Tray not ejected from BD Rhapsody™ Scanner

Possible causes	Recommended solutions
Cartridge in incorrect orientation or no cartridge	Tap the eject button at the right top corner of the cartridge insertion screen. Reinsert the cartridge on the tray.

BD Rhapsody Scanner not analyzing all images

Possible causes	Recommended solutions
Masked or incompletely processed wells	The software automatically corrects for masked or incompletely processed wells. No manual extrapolation is necessary.

Persistent scanning artifacts

Artifacts might include apparent cells in an empty hemocytometer, consistently low capture rate warnings, auto-exposure errors, and dirty optics cover.

Possible causes	Recommended solutions
Dirty optics	Contact BD Biosciences technical support at researchapplications@bd.com .

Cartridge loading troubleshooting

Introduction

This topic describes possible problems and recommended solutions for BD Rhapsody™ Cartridge issues. These issues arise during image analysis. Also see [BD Rhapsody™ Scanner metrics \(page 119\)](#).

Number of cells captured too high or cell doublet rate too high (out of range)

Possible causes	Recommended solutions
Too many cells loaded	<ul style="list-style-type: none"> • Confirm that the calculation of cell dilution is correct. • Reduce the number of cells loaded in the cartridge.
Improper counting with hemocytometer	<ul style="list-style-type: none"> • Recount cells with the BD Rhapsody Scanner. • Follow Processing cells with the BD Rhapsody™ Single-Cell Analysis system (page 65).
Incorrect calculation of cell concentration	<ul style="list-style-type: none"> • Use the BD Rhapsody Scanner for cell counting and analysis. • Use the Analysis app Sample Calculator for preparing cell suspensions.

Number of cells captured lower than anticipated

Possible causes	Recommended solutions
Incorrect calculation of cell concentration	<ul style="list-style-type: none"> Use the BD Rhapsody Scanner for cell counting and analysis. Use the Analysis app Sample Calculator for preparing cell suspensions.
Cell size larger than recommended range	Cell capture efficiency is reduced in the BD Rhapsody Cartridge if the cell diameter is >20 μm .

Bead loading density out of range

Possible causes	Recommended solutions
Insufficient number of Cell Capture Beads loaded	<ul style="list-style-type: none"> Ensure that all of the beads are pelleted before removing storage buffer and are resuspended in Sample Buffer (Cat. No. 650000062). Use the Bead Load mode on the pipette. Ensure the beads are well suspended before loading into the cartridge.

Image analysis fails at Bead Wash, indicating insufficient removal of excess Cell Capture Beads

Possible causes	Recommended solutions
Pipette mode incorrect	Ensure that the correct pipette mode is used for the step.
Pipette step missed	Review protocol, and repeat, if necessary.

Cell retention out of range, indicating cell loss during cartridge workflow

Possible causes	Recommended solutions
Pipette mode incorrect	Ensure that the correct pipette mode is used for the step.
Cell viability low	Use cells of high viability.

Bead retrieval efficiency out of range, indicating poor retrieval

Possible causes	Recommended solutions
Retrieval (top) magnet not positioned properly	Check magnet position.
Pipette mode incorrect	Ensure that the correct pipette mode is used for the step.

Image analysis does not start

Possible causes	Recommended solutions
Software does not initiate analysis after a scan	<ol style="list-style-type: none"> 1. On the BD Rhapsody Scanner main menu, tap Analysis. 2. Tap the experiment. 3. Tap Analyze to start the analysis. You can analyze multiple scan steps at the same time.

Dropped the cartridge or hit it against object

Possible causes	Recommended solutions
Various	<ul style="list-style-type: none"> • If the cartridge was dropped, BD recommends using a new cartridge; otherwise, carefully review the imaging metrics before proceeding. • If the cartridge was struck, proceed at your own risk, and carefully review the imaging metrics before proceeding.

Air bubble in cartridge

Possible causes	Recommended solutions
Air bubble present in pipette tip while dispensing buffer	<p>Confirm that there is an air bubble in the cartridge by examining an image of the cartridge.</p> <p>Ensure that the pipette tip contains only buffer and no air bubble is trapped in the cartridge at the end of aspiration.</p>
Re-used pipette tip	Use a new pipette tip at every pipetting step.

Image analysis fails Contact BD Biosciences technical support at researchapplications@bd.com.

Installation or mechanical issues See the *BD™ Rhapsody Single-Cell Analysis System Installation and Maintenance Guide* (Doc ID: 43084).

BD Rhapsody Scanner software messages

Introduction

This topic lists error and warning messages that you might encounter while using the BD Rhapsody Scanner software.

If the recommended solutions do not resolve the problem, contact BD Biosciences tech support at researchapplications@bd.com or 1.877.232.8995, prompt 2, 2.

Error messages

Error messages require you to perform an action, according to a displayed dialog, before you can proceed.

Error message	Possible causes	Recommended solutions
“Failed to open door. Ensure nothing is blocking the door.”	Obstruction	Remove obstruction, and retry opening the scanner cartridge door.
“Failed to open door.”	Various; not an obstruction	Restart scanner.
“Failed to close door. Ensure nothing is blocking the door.”	Obstruction	Remove obstruction, and retry closing the scanner cartridge door.
“Failed to close door.”	Various; not an obstruction	Restart the scanner.
“Obstruction detected while ejecting.”	Obstruction or calibration update required	Remove obstruction, and retry ejecting the cartridge.
“Obstruction detected while retracting stage.”	Obstruction	<ol style="list-style-type: none"> 1. Remove obstruction, and retry retracting cartridge into the scanner. 2. Click Cancel. 3. Restart the scanner.

Error message (continued)	Possible causes	Recommended solutions
“Available disk space is extremely low.”	Low disk space	Contact BD tech support researchapplications@bd.com or 1.877.232.8995, prompt 2, 2.
“Failed to initialize...”	Various	Contact BD tech support researchapplications@bd.com or 1.877.232.8995, prompt 2, 2.
“Requested move not safe.”	Various	Contact BD tech support researchapplications@bd.com or 1.877.232.8995, prompt 2, 2.
“Unknown cartridge barcode.”	Unsupported barcode scanned	Contact BD tech support researchapplications@bd.com or 1.877.232.8995, prompt 2, 2.
“Failed to read file.”	Various	Contact BD tech support researchapplications@bd.com or 1.877.232.8995, prompt 2, 2.
“Cartridge alignment feature not found.”	<ul style="list-style-type: none"> ● Incorrect cartridge ● Light failed 	Rescan the cartridge.

Warnings

Warnings display in the Warnings box on the scanner display and are mostly informative messages that require no action from you.

Warning message	Possible causes	Recommended solutions
“Barcode scan failed. Please make sure the cartridge is inserted and aligned correctly.”	Barcode not read	<ol style="list-style-type: none"> 1. Reinsert the cartridge, and rescan the cartridge. 2. If necessary, restart the scanner.
“Available disk space is running low.”	Limited disk space	Contact BD tech support researchapplications@bd.com or 1.877.232.8995, prompt 2, 2.
“The protocol was not found.”	Protocol missing	Contact BD tech support researchapplications@bd.com or 1.877.232.8995, prompt 2, 2.
“Auto-exposure is not configured for image type. Using default value instead.”	Method not found	Contact BD tech support researchapplications@bd.com or 1.877.232.8995, prompt 2, 2.
“Auto-exposure resulted in images darker than normal.”	<ul style="list-style-type: none"> • No cell in image tile • Light failed 	Contact BD tech support researchapplications@bd.com or 1.877.232.8995, prompt 2, 2.
“Auto-exposure resulted in images brighter than normal.”	Various	Contact BD tech support researchapplications@bd.com or 1.877.232.8995, prompt 2, 2.
“Auto-exposure adjustment unsuccessful. Using nearest value.”	Algorithm calculation	Contact BD tech support researchapplications@bd.com or 1.877.232.8995, prompt 2, 2.

Warning message (continued)	Possible causes	Recommended solutions
“Autofocus did not find perfect focus. Using nearest value.”	<ul style="list-style-type: none"> ● Bubble in cartridge ● Calibration needs update 	Contact BD tech support researchapplications@bd.com or 1.877.232.8995, prompt 2, 2.
“Attempted to use exposure time greater than maximum. Using the maximum instead.”	Light failed	Contact BD tech support researchapplications@bd.com or 1.877.232.8995, prompt 2, 2.
“No alignment marks found. Continuing without cartridge specific calibration.”	Cartridge misaligned	Reinsert cartridge, and rescan.
“Not enough local alignment marks detected.”	<ul style="list-style-type: none"> ● Bead is covering an alignment mark ● Bad image 	Contact BD tech support researchapplications@bd.com or 1.877.232.8995, prompt 2, 2.

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A

BD Rhapsody™ Scanner metrics

- Image analysis metrics (page 120)
- Reviewing the analysis metrics (page 124)

Image analysis metrics

Understanding analysis metrics The chart lists the steps and metrics measured during image analysis of the hemocytometer or BD Rhapsody™ Cartridge.

Step	Metric	Definition	Use	Passing threshold
Hemocytometer	Total cell concentration (cell/ μ L)	Number of viable (Calcein AM-positive) and non-viable (DRAQ7-positive) cells/ μ L	Calculates concentration of the cell suspension to determine the volume of cell suspension to use for loading onto the cartridge.	Value
Hemocytometer	Cell viability (%)	Percent viable (Calcein AM-positive) cells	Assesses cell viability to determine if the cell sample meets the quality threshold.	Value
Cell Load	Number of wells with viable cells at Cell Load	Number of wells containing viable (Calcein AM-positive) cells	Provides a preliminary estimate of the number of wells with viable cells captured. ^a	—
Cell Load	Number of viable cells captured in wells at Cell Load	Number of viable (Calcein AM-positive) cells captured in a well	Provides a preliminary estimate of the total number of viable cells captured in the cartridge. ^b	—

Step (continued)	Metric	Definition	Use	Passing threshold
Cell Load	Cell multiplet rate at Cell Load ^d (%)	Number of wells containing multiple viable (Calcein AM-positive) cells versus the total number of wells containing one or more viable cells	Provides a measure of cell clumping.	—
Bead Wash	Number of wells with viable cells and a bead	Total number of wells with one bead and ≥ 1 viable (Calcein AM-positive) cells	Provides an estimate of the number of wells with viable cells captured with beads. ^c	Value
Bead Wash	Number of viable cells captured in wells with a bead	Total number of viable (Calcein AM-positive) cells captured in a well with a bead	Estimates the total number of viable cells captured on a Cell Capture Bead at cell lysis. ^c	—
Bead Wash	Cell multiplet rate ^d (%)	Number of wells containing one bead and multiple viable (Calcein AM-positive) cells versus the total number of wells containing one bead and one or more viable cells	Provides an estimate of the occurrence of multiple cells captured by the same bead in a well.	Value

Step (continued)	Metric	Definition	Use	Passing threshold
Bead Wash	Bead loading efficiency (%)	Percent wells with one bead	Indicates if the cartridge is significantly underloaded with beads.	≥80%
Bead Wash	Cell retention rate (%)	Number of viable (Calcein AM-positive) cells captured in wells prior to lysis versus number of viable cells captured in wells prior to bead loading	Indicates if a significant number of cells initially loaded into wells are lost or died during the workflow prior to lysis.	≥60%
Retrieval	Bead retrieval efficiency (%)	Percent of wells with beads removed	Indicates if the number of beads retrieved is significantly lower than expected.	≥90%

- a. The number of wells with viable cells is less than the total number of viable cells captured in wells because of wells containing cell multiplets.
- b. Cell capture efficiency at Cell Load (%) = $\frac{\text{Number of viable cells captured in well at Cell Load}}{\text{Total number of viable cells imaged}} * 100$.
- c. The metric, the number of wells with viable cells and a bead, is less than the metric, the number of viable cells captured in wells with a bead, due to wells containing cell multiplets. Multiple cells captured with one bead will appear as one cell in sequencing data. Because of this, the appropriate metric to use for an estimation of the number of cells that could be recovered in sequencing is the metric, number of wells with viable cells and a bead.
- d. The estimated multiplet rate is calculated by Poisson distribution using the number of cells loaded into the cartridge. See [Processing cells with the BD Rhapsody™ Single-Cell Analysis system \(page 65\)](#). The cell multiplet rate reported by the BD Rhapsody Scanner is determined by imaging. If the cell multiplet rate >> estimated multiplet rate, this might indicate cell clumping.

**Viewing image
analysis metrics**

To view image analysis metrics in a spreadsheet, click the **Rhapsody Data** shortcut on the BD Rhapsody Scanner screen. The Rhapsody Data folder contains ExperimentName_SampleName.csv, PrepareSampleResults.csv, and Cell Count.csv.

Reviewing the analysis metrics

1. Navigate to the Analysis application.
2. Tap the experiment to ensure that the analyses are completed.
For example:

Dispense1-1

Cartridge 0109034055A

Scan Date	Sample	Step	Analysis Status
2018-02-21 10:29:27.56	Dispense1-1	Cell Load	✓ Completed
2018-02-21 10:57:04.44	Dispense1-1	Bead Load	✓ Completed
2018-02-21 11:11:49.59	Dispense1-1	Bead Wash	✓ Completed
2018-02-21 11:26:42.94	Dispense1-1	Retrieval	✓ Completed

3. Review the analysis metrics. For PASS criteria, see [Image analysis metrics \(page 120\)](#). For example:

Analysis	
Number of wells with viable cells at cell load	9118
Cell multiplet rate at cell load	2.4 %
Number of wells with viable cells and a bead	7999
Cell multiplet rate	2.0 %
Bead loading efficiency	✓ PASS
Excess bead rate	✓ PASS
Cell retention rate	✓ PASS
Bead retrieval efficiency	✓ PASS

4. (Optional) View quantitative analysis metrics for each experiment in a spreadsheet. For example, the .csv files from the analysis of the Jurkat and Ramos cells are displayed:

Note: To set up the shortcut to the Rhapsody Data folder in Quick Access, see [BD Rhapsody Scanner software \(page 29\)](#).

- (1) [Analysis metrics \(page 126\)](#)
- (2) [Cell count \(page 127\)](#)
- (3) [Sample result \(page 127\)](#)

(1) Analysis metrics

Barcode	0109034055A
Experiment	20180221_SL_XH
Sample	Dispense1-1
Scan System ID	R*****-1017
Scan start date and time	2/21/2018 10:29
Operator	SL
Cell load analysis last updated	2/21/2018 10:34
Total number of tiles	26
Cell load number of tiles processed	26
Number of wells with viable cells at cell load	9118
Number of viable cells captured in wells at cell load	9344
Cell multiplet rate at cell load	2.40%
Bead wash analysis last updated	2/21/2018 11:20
Bead wash number of tiles processed	26
Number of wells with viable cells and a bead	7999
Number of viable cells captured in wells with a bead	8162
Cell multiplet rate	2.00%
Bead loading efficiency	94.40%
Bead loading acceptance	PASS
Excess bead rate	0.00%
Excess bead acceptance	PASS
Cell retention rate	90.20%
Cell retention acceptance	PASS
Bead retrieval analysis last updated	2/21/2018 11:37
Bead retrieval number of tiles processed	26
Bead retrieval efficiency	94.80%
Bead retrieval acceptance	PASS

(2) Cell count

Analysis software version	1.0.8.7
Experiment	20180221_SL_XH
Sample	Ramos
Barcode	0107001030A
Protocol step	Side B
Scan System ID	R*****-1017
Operator	SL
Scan start date and time	2/21/2018 10:06
Analysis last updated	2/21/2018 10:06
Total cell concentration (live and dead)	331.67
Cell viability	91.87

(3) Sample result

Experiment	20180221	SL_XH					
Cartridge type	109						
Cartridge volume (uL)	575						
Total volume to prepare (uL)	3000						
Desired number of captured cells	10000						
Sample buffer volume (uL)	2816.8						
Loading cell concentration	19.9						
Cell doublet rate (%)	2.4						
Sample	Operator	Scan Date and Time	Concentration	Viable Cells	Relative Amount	Stock Volume (uL)	
Jurkat	SL	06:09.4	320.2	50.63	1	93.2	
Ramos	SL	06:28.8	331.7	91.87	1	90	

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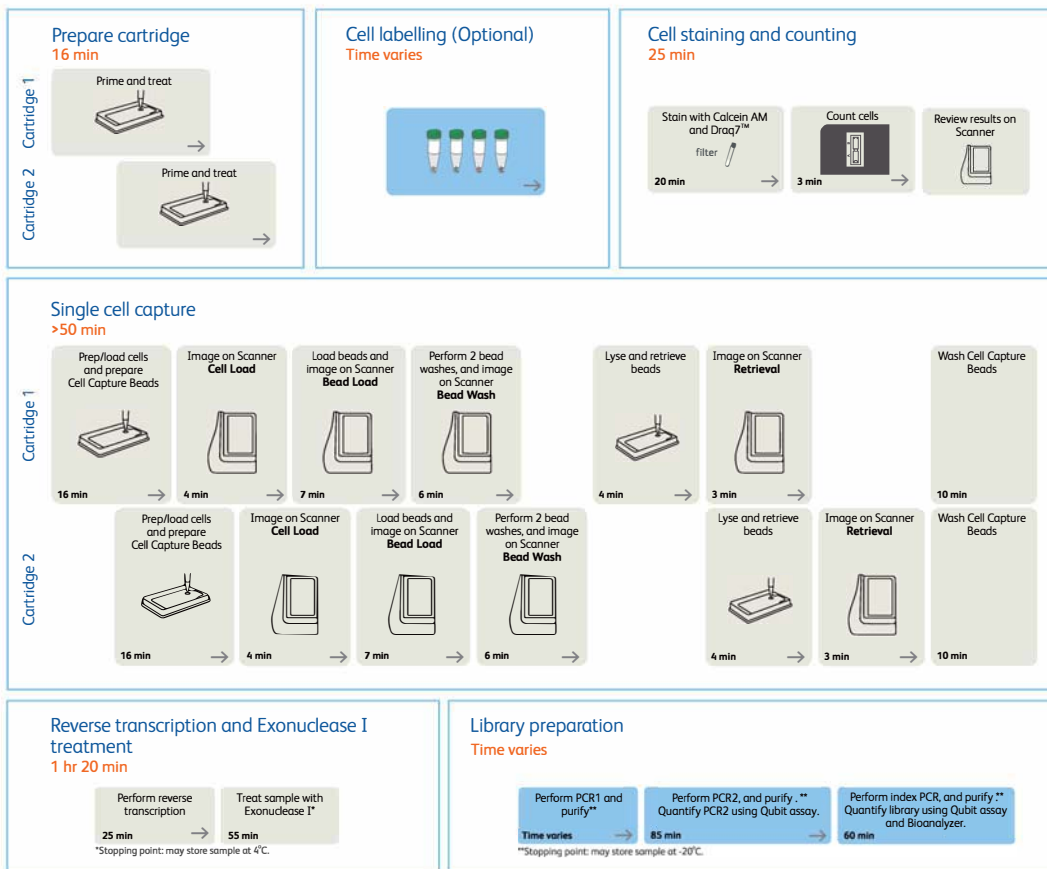
B

Workflow with two BD Rhapsody™ Cartridges

- Workflow with two cartridges (page 130)
- Best practices with a two-cartridge workflow (page 131)

Workflow with two cartridges

Staggered boxes indicate staggering the start of like steps.



Best practices with a two-cartridge workflow

Reagent preparation

To prepare a master mix of sufficient volume for two cartridges, follow the volumes for two libraries plus 10% overage listed for preparing a master mix.

To perform the workflow, follow the *Single Cell Analysis Workflow with BD Rhapsody™ Systems* (Doc ID: 220524).

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C

Updating the BD Rhapsody™ Scanner software

- Introduction (page 134)
- Uninstalling scanner software (page 134)
- Installing scanner software (page 135)
- Launching the scanner software (page 138)

Introduction

BD Biosciences recommends updating the scanner software by navigating the installation with keyboard and mouse. For recommended peripherals, see a BD Rhapsody™ user guide.

This appendix provides information for:

- [Uninstalling scanner software \(page 134\)](#)
 - [Installing scanner software \(page 135\)](#)
 - [Launching the scanner software \(page 138\)](#)
-

Uninstalling scanner software

Procedure

If BD Rhapsody™ Scanner software v1.0.8 or earlier is installed, follow this procedure. If the installed version is later than v1.0.8, proceed to [Installing scanner software \(page 135\)](#).

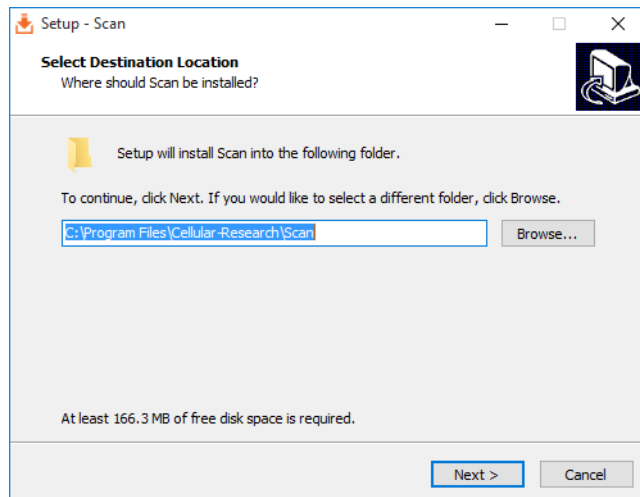
1. On the scanner desktop, click **Start** in the lower left corner, and then click **Settings**. The Settings window is displayed.
2. Click **System**. A System window is displayed.
3. Select the **Apps and features** menu item.
4. In the **Apps and features** pane, select **Rhapsody Scanner - Scan 1.0.8**.
5. Click **Uninstall**, and click **Uninstall** again in the warning dialog. A User Account Control dialog is displayed.
6. Click **Yes**, and click **Yes** again in the **Scan Uninstall** warning dialog.
7. After the uninstall task is complete, click **OK**.

- Repeat steps 4–7 to uninstall **Select Rhapsody Scanner - Analysis 1.0.8**.
 - Close the **Settings** window.
-

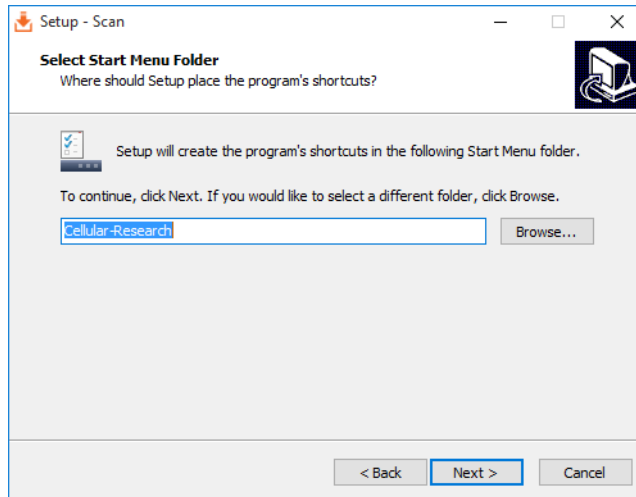
Installing scanner software

For successful installation of the software, install the Scan software before installing the Analysis software. If you install the Analysis software first, uninstall it, and then install the Scan software.

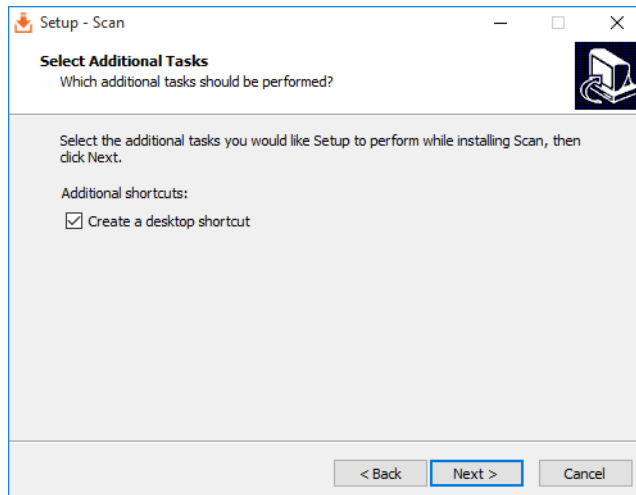
- Insert the USB drive with the scanner software into one of the ports of the scanner.
- In Microsoft® Windows®, Open a File Explorer window, and navigate to the USB drive directory.
- Double-click **Scan 1.1.0.2 Setup.exe** or later. A User Account Control dialog is displayed.
- Click **Yes**. The Select Destination Location dialog is displayed.



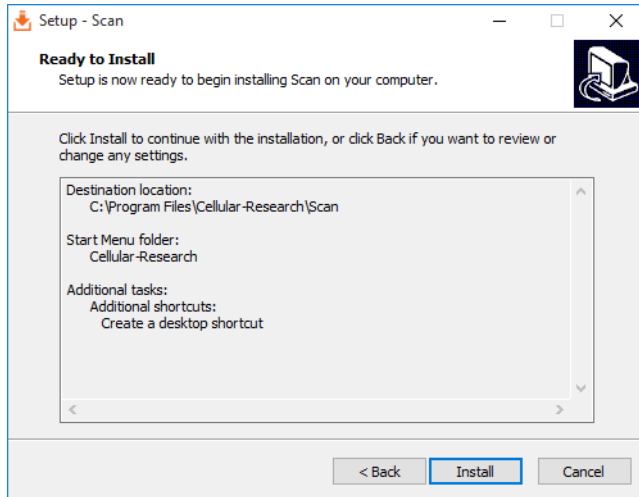
5. Click **Next**. The Select Start Menu Folder dialog is displayed.



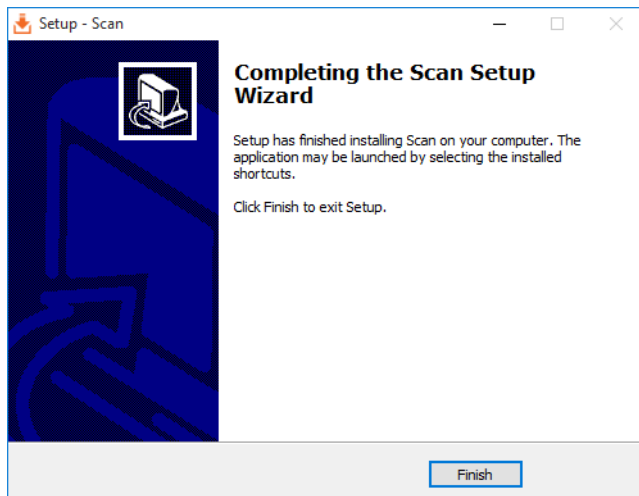
6. Click **Next**. The Select Additional Tasks dialog is displayed.



7. Select the **Create a desktop shortcut** checkbox, and then click **Next**. The Ready to Install dialog is displayed.



8. Click **Install**. The Completing the Scan Setup Wizard window is displayed.



9. Click **Finish**.

10. Repeat steps 1–9 to install **Analysis 1.1.0.2 Setup.exe** or later.
 11. Remove the USB drive from the scanner, and store it.
-

Launching the scanner software

1. On the scanner desktop, double-click **Scan**. The scanner software home page is displayed.
 2. Tap **About**, and confirm the scanner software version.
-

D

Safety

- General safety and limitations (page 140)
- Chemical safety (page 140)
- Physical safety (page 142)
- Instrument waste disposal (page 142)

General safety and limitations

For instrument safety, see the *BD Rhapsody™ Express Instrument and BD Rhapsody™ Scanner Safety and Limitations Guide* (Doc ID: 42061).

Genomics technical publications are available for download from the BD Genomics Resource Library at bd.com/genomics-resources.

Chemical safety

Requirements

- Read and comprehend all safety data sheets (SDSs) by chemical manufacturers before you use, store, or handle any chemicals or hazardous materials.
 - Wear personal protective equipment (gloves, safety glasses, fully enclosed shoes, lab coats) when handling chemicals.
 - Do not inhale fumes from chemicals. Use adequate ventilation, and return caps to bottles immediately after use.
 - Check regularly for chemical spills or leaks. Follow SDS recommendations for cleaning up spills or leaks.
-

Waste

The BD Rhapsody™ system has two waste types or streams. Each waste stream requires individual consideration for safe and responsible disposal:

Waste	Description
Stream 1: Waste Collection Container	<ul style="list-style-type: none">● Frequency of Handling: every BD Rhapsody™ experiment● Content: ethanol (11%), polymer microparticles (<1%), cells (trace)● Main Risk Constituent: cells (trace)● Collect and dispose of all waste in the Waste Collection Container using proper precautions and according to local safety regulations.
Stream 2: BD Rhapsody™ Cartridge	<ul style="list-style-type: none">● Frequency of Handling: every BD Rhapsody experiment● Content: polymer (99%), polymer microparticles (<1%), lysis buffer (<1%)● Main Risk Constituent(s): lysis buffer● Collect and dispose of all used BD Rhapsody™ Cartridges using proper precautions and according to local safety regulations.

Physical safety

See the *BD Rhapsody™ Express Instrument and BD Rhapsody™ Scanner Safety and Limitations Guide* (Doc ID: 42061).

Genomics technical publications are available for download from the BD Genomics Resource Library at bd.com/genomics-resources.

Instrument waste disposal

Disposal of the instrument

Contact BD Biosciences technical support at researchapplications@bd.com before moving the BD Rhapsody Scanner or disposing of the BD Rhapsody™ Express instrument or the BD Rhapsody Scanner. For more information, see [Instrument technical support \(page 13\)](#).

Glossary

B

BD Rhapsody™ Express instrument Mechanical station used for loading Cell Capture Beads and cells into the BD Rhapsody™ Cartridge.

BD Rhapsody™ Scanner Instrument used for counting cells on a hemocytometer and counting viable cells in the cartridge workflow.

BD Rhapsody™ Single-Cell Analysis system The system includes:

- BD Rhapsody Express instrument
- BD Rhapsody™ P1200M pipette
- BD Rhapsody™ P5000M pipette
- BD Rhapsody Scanner
- Hemocytometer Adapter
- Bioinformatics software and sequencing analysis

L

load To add a reagent to the BD Rhapsody Cartridge.

P

protocol The protocol files for the BD Rhapsody Scanner describe all instrument settings for a cartridge scan and the order in which the scans occur.

S

scan A series of images taken of the Hemocytometer Adapter or BD Rhapsody Cartridge by the BD Rhapsody Scanner at multiple predetermined locations.
